

Annual Research & Review in Biology 8(4): 1-13, 2015, Article no.ARRB.20527 ISSN: 2347-565X, NLM ID: 101632869



SCIENCEDOMAIN international www.sciencedomain.org

European Sea Bass (*Dicentrarchus labrax* L. 1758) As a Sentinel Species in Europe to Study the Effects of Contaminants

Daniela Conti^{1*}, Stefania Balzamo¹, Andrea Paina¹, Cristina Martone¹, Elisa Raso¹, Fabio Cadoni¹, Federica Savorelli², Manorama Croppo², Vanessa Bellaria¹ and Alessandra Pati¹

¹ISPRA - Institute for Environmental Protection and Research, Environmental Metrology Unit, Via di Castel Romano 100 – 00128, Rome, Italy. ²ARPA - Agenzia Regionale Prevenzione e Ambiente dell'Emilia Romagna, Sezione Provinciale di Ferrara, via Bologna 534 – 44124, Ferrara, Italy.

Authors' contributions

This work was carried out in collaboration between all authors. Authors SB and DC planned and organized all the experimental activities of the project "Inclusion of the European sea bass (Dicentrarchus labrax L. 1758) among species recommended for testing in the OECD Guidelines for testing of chemicals". Authors DC, AP, CM, ER, FC, FS, MC, VB and AP performed the acute toxicity tests with sea bass. Authors DC and SB wrote the manuscript. Authors AP, FS, MC and CM contributed to the paper preparation. All authors read and approved the final manuscript

Article Information

DOI: 10.9734/ARRB/2015/20527 <u>Editor(s)</u>: (1) George Perry, Dean and Professor of Biology, University of Texas at San Antonio, USA. <u>Reviewers</u>: (1) Jairo Pinheiro Da Silva, Federal Rural University of Rio De Janeiro, Brazil. (2) André Lincoln Barroso Magalhães, Federal University of São João del-Rei, Brazil. Complete Peer review History: <u>http://sciencedomain.org/review-history/11775</u>

Mini-review Article

Received 30th July 2015 Accepted 20th September 2015 Published 9th October 2015

ABSTRACT

Aim: The current study was designed to evaluate whether European Sea Bass (*Dicentrarchus labrax* L. 1758) can be used as a sentinel species to test contaminants. For this aim the acute lethality (96 h) of anionic surfactant sodium dodecyl sulfate (SDS) on sea bass juveniles at two different water salinity percentages (20‰ and 5 ‰) were tested in order to assess how the water salinity may affect the sensitivity of the sea bass to SDS. A comparison with results from scientific literature especially freshwater species was also reported.

*Corresponding author: E-mail: stefania.balzamo@isprambiente.it;

Study Design: Five different sized fish groups (from 0.068 g to 1.42 g) were assessed in 96-h semi-static tests according to OECD guideline n. 203 (1992). The average 96h-LC₅₀ (plus 95% confidence interval) values were calculated using SDS nominal concentrations.

Results: At a percentage of salinity of 20‰, the 96h-LC₅₀ values increased from 5.76 mg/l to 9.50 mg/l increasing the size of juveniles as well. SDS at a salinity of 5‰, was always found to be significantly less toxic (96h-LC₅₀: from 13.50 mg/l to 14.87 mg/l) independently of fish size.

Conclusion: The results of this study show that the characteristics of *D. labrax* make this species useful to provide information of toxicity in both seawater and freshwater. Its ease of maintenance and testing under laboratory conditions, and its broad euryhalinity confirm the convenience of use it as a sentinel species for detecting environmental impact and as a rule in monitoring studies. A comparison with other Italian and international data on sea bass is reported.

Keywords: Anionic surfactant; SDS; acute toxicity; European sea bass (Dicentrarchus labrax); salinity.

1. INTRODUCTION

Sea bass is a perciform teleost fish belonging to the Moronidae family and the genus *Dicentrarchus*

(<u>http://researcharchive.calacademy.org/research/</u> ichthyology/catalog/fishcatmain.asp).

The species *Dicentrarchus labrax* L. 1758, named European or common sea bass, is primarily a marine fish, but it is also found in brackish and freshwater [1]. It was the first marine non-salmonid species to be commercially cultured in Europe and, actually, it is the one of the most important commercial fish widely reared in the Mediterranean areas: Greece, Turkey, Italy, Spain, Croatia and Egypt are the biggest producers [2].

Usually the adult sea bass is a highly eurythermal and euryhaline species. In Winter, it lives in waters at 5-6°C in the lagoons of the northern Mediterranean. In Summer, it is abundant in waters where temperatures exceed 30°C. *D. labrax* can also survive in a wide variety of environmental salinity levels (0.5 - 35%) [3,4].

It is well known that metabolism, development and growth of teleost fishes are controlled by internal factors (CNS, endocrinological and neuroendocrinological systems), but they are also highly dependent on environmental conditions [5]. Many studies have reported that water salinity is one of the determining factors (together with temperature and photoperiod) which directly acts through receptors to increase or decrease the growth and the development of fish [6].

For the purpose of this study, *D. labrax* was selected because of its ease of culture and maintenance under laboratory conditions. It is a well-known species, frequently used to

investigate the toxicity of various classes of chemicals [6-18] and also acts as a sentinel species (i.e. indicator species or bioindicator species) on monitoring studies [19-22].

Dicentrarchus labrax has extensively been used in Italy for regulatory evaluation of ecotoxicity. At least three important national projects concerning the sea bass were carried out. The first entitled "Priority Substances: Fish (D. labrax, Cyprinus carpio) as target organisms in toxicity, bioaccumulation and genotoxicity tests". It aimed at the toxicological assessment of dangerous recommended chemicals by the Italian Legislative Decree 152/2006 (Italian transposition of WFD 2000/60/CE) [23]. The second project entitled "Oil dispersants: toxicity testing with marine organisms (Algae, Crustaceans and Fish)". The fish species selected in this study (D. labrax and Sparus aurata) were included in the Italian Decree of the Environment Ministry (25 February 2011, Official Journal n. 74, 31 March 2011 - Supplement n. 87) concerning the procedures for assessing suitability of oil absorbent and dispersant products to be employed in marine environment, in order to minimize the impact of oil spills [24].

The latter project organized by UNICHIM (a National no-profit organization associated to the Italian Authority for Standardization, UNI) aimed to validate the guideline n° 203 of the Organisation for Economic Co-operation and Development (OECD) by using *D. Labrax* [25,26]

Furthermore, in Italy, the sea bass at different life stages (eggs, larvae, post-larvae and juveniles) was preferably used for testing various classes of chemicals [27-30].

Recently, a proposal concerning the "Inclusion of the European sea bass (*D. labrax* L.1758) among species recommended for testing in the

OECD Guidelines for testing of chemicals n° 203, 212, 215, 305" was presented by Italy at the 25th Meeting of the Working Group of National Coordinators of the Test Guidelines Programme (WNT 25) held from 9 to 11 April 2013. OECD Members highlighted the lack of representativeness of fish species in the toxicity testing guidelines (TG) as far as the Mediterranean Sea is concerned. They proposed to Italy to develop a Guidance Document (GD) on the use of the European sea bass in fish toxicity and bioaccumulation tests.

The quality of test organism is key to successful conduct of fish test and species selection should consider a number of different criteria including: size, ease of maintenance, convenience for relevant economic biological testing. or ecological factors, known sensitivity, pre-existing data, animal welfare, availability of cultured, as opposed, to field-collected organisms as well as regional preferences national or [31]. Dicentrarchus labrax's compliance to most of these criteria was clearly recognized. The main strengths for using this species are its wide availability at every stage of the reproductive cycle in many countries of the Mediterranean Basin (Italy, Spain, Turkey, Greece, France and North Africa), its ease of maintenance and testing under laboratory conditions, and its broad eurvhalinity. On the other hand, there are not any recommended estuarine and marine fish species in the acute (OECD TG 203), prolonged (OECD TG 204) [32] and juvenile growth test (OECD TG 215) [33]. Nevertheless, the OECD test guidelines present no restrictions in the choice of fish test species, provided that the alternative species fulfils the selection criteria mentioned above and the rationale for the choice is well documented.

Surfactants are economically important chemicals which are widely used in household cleaning detergents, personal care products, textiles, paints, pharmaceuticals, cosmetics, etc. They and their products are often discharged directly to waters or into sewage treatment plants and then dispersed into the environment. Due to their widespread use and their presence in the environment, a lot of studies were carried out to demonstrate that surfactants are capable of causing important adverse effects to the aquatic and terrestrial life [34].

Surfactants are amphiphilic compounds capable of reducing the surface tension of water by forming a monolayer at the air-water interface. They are water-soluble in the micro- to millimolar range above which they self-associate to micelles, bilayer vesicles or other aggregates [35].

The sodium dodecyl sulfate ($C_{12}H_{25}NaO_4S$ - SDS), also known as sodium lauryl sulfate, belongs to the class of anionic surfactants (fatty alcohol sulphates, AS). SDS exerts a low logP (partition coefficient octanol/water) and logHCl (partition coefficient water/air) value and is miscible in water (water solubility is 100g/l). Because of this, it can be easily handled and hampering sorption to exposure vessels or evaporation can be excluded [36].

The current study was designed to evaluate the acute lethality (96h) of anionic surfactant SDS on sea bass juveniles. Two different water salinity percentages (20‰ and 5‰) were tested in order to assess how the water salinity may affect the sensitivity of the sea bass to SDS. A comparison with SDS results from scientific literature was also reported.

2. MATERIALS AND METHODS

2.1 Test Chemical

Sodium Dodecyl Sulfate (CAS N° 151-21-3; EINECS: 205-788-1) Panreac PA-ACS (99% purity) was purchased from Nova Chimica (batch n° 367414).

2.2 Test Fish

Dicentrarchus labrax juveniles were supplied by two different salt water fish farms: Ca' Zuliani Lagoon Fishing Farm (Porto Tolle, Rovigo -Veneto region, northern Italy) and Caldoli Fishing Farm (Lesina, Foggia - Puglia region, southern Italy). All fishes were acclimated for 14 days to laboratory conditions in breeding tanks equipped with mechanical and biological filter and a recirculating pump with capacity of 1000 liter/hour at the same conditions of the origin fishing farm. Breeding tanks (300 liter capacity) were filled with artificial seawater, fish loading g/l. was approximately 0.5-0.6 Water temperature had been keeping at 20±1℃ in continuous artificial aeration; oxygen concentration was between 7 to 9 mg/l and pH ranged from 7.2 to 8.5. After 48h we started to check the mortality for seven days; the mortality was less than 5% then the batch was accepted for testing. The juveniles sea bass were adapted

to salinity variation gradually in a time of 14 days (salt concentration was measured with Atago® salinometer, accuracy 0.1%). The fishes were regularly fed (about 2% by fish weight) with commercial fish food provided by the farmers with a suitable composition to their nutritional needs.

The concentration of the main catabolites, NH_4 , NO_2 and NO_3 (Salifert – Ammonia, Nitrite and Nitrate Prof-Test kit), the dissolved oxygen concentration (HI9143 portable Dissolved Oxygen Meter, Hanna Instruments) and the pH value of the water (HI9025C portable Waterproof pH meters, Hanna Instruments) were measured every day. Ammonia and nitrite are usually absent in the breeding water (nitrate concentration is acceptable until 30 mg/l).

The fish specimens were divided in five different groups according to their size; the mean values of weight (g) and total length (cm) are shown in Table 1.

The juvenile sea bass specimens have been led from 5‰ to 20‰ of salt water gradually at least fifteen days before they are used.

2.3 Acute Toxicity Tests

European sea bass, *D. labrax*, were tested in a 96-h semistatic test according to OECD guideline n° 203 (1992). Seven independent toxicity tests

were conducted in a room with controlled temperature and photoperiod (16hL:8hD). Based on the fish sizes, different test chambers were used (test organism loading not exceeding 1 g/l) in the tests. The quality of water was monitored on a daily basis for the duration of the tests by measuring water dissolved oxygen, pH and temperature. No food was provided to fish during the tests.

The nominal exposure concentrations used in the SDS definitive assays were selected according to a preliminary range-finding test, as they are shown in Table 2.

Nominal SDS concentrations were obtained by serial dilutions of a stock solution (100 mg/l) prepared in ultrapure water immediately before use.

The fatty alcohol sulphates such as SDS, are among the most rapidly biodegradable compounds in the environment. Steber et al., 1988 [37] reported for fatty alcohol sulphates a primary degradation of 95-98% in a 5 days period. SDS biodegradation was reported in Antarctic coastal waters with half lives of 160-460 hours [38]. In order to minimize disturbance of the test organisms and the degradation of SDS, test solutions were carefully renewed every 24 or 48h. A saltwater control was included in each experiment.

 Table 1. Features of fishes Dicentrarchus labrax (Moronidae) used in experimental procedures (in increasing order)

Fish group	N° of specimens	W(g)	TL(cm)	Fishing farm
	150	0.068±0.03	2.23±0.22	Ca Zuliani Lagon Fishing Farm, Rovigo, Italy
	150	0.50±0.07	3.90±0.30	Caldoli Fishing Farm, Foggia, Italy
	140	0.66±0.22	4.64±0.29	Caldoli Fishing Farm, Foggia, Italy
IV	140	0.94±0.34	4.79±0.52	Ca Zuliani Lagon Fishing Farm, Rovigo, Italy
V	140	1.42±0.05	5.40±0.07	Caldoli Fishing Farm, Foggia, Italy

Abbreviations: W: weight; TL: total length

Table 2. Exposure conditions used in the sodium dodecyl sulfate acute toxicity testing on Dicentrarchus labrax (Moronidae)

Expt	Fish group	Salinity (‰)	T (°C)	рН	Renewal	n°org/ chamber	R	Total n° fish	SDS nominal concentrations (mg/l)
1	1	20	20.0±0.7	8.5±0.1	every 48h	7	3	126	2.5 - 3.6 - 5.0 -7.1-10
2	11	5	19.6±0.3	8.4±0.2	every 48h	7	3	126	10 -16 - 25 - 40 - 63
3	111	20	19.5±1	8.4±0.2	every 48h	10	1	60	1 - 5 - 9 -13 -17
4		5	19.5±1	8.4±0.2	every 48h	10	1	60	5 - 9 -13 -17- 21
5	IV	20	20.0±1	8.5±0.1	every 48h	7	3	126	2.5 - 5 -10 - 20 - 40
6	V	5	20.5±1	8.5±0.1	every 24h	10	1	60	6 - 9 - 12 -15 -18
7	V	20	20.5±1	8.4±0.1	every 24h	10	1	60	3.5 - 5.5 - 7.5- 9.5 -11.5

Abbreviations: R: replicate

A summary of the exposure conditions used in each experiment is shown in Table 2.

Fish were inspected every 30 minutes for the first 8 hours and subsequently at 24, 48, 72 and 96h for mortality and other visible abnormalities (e.g. swimming behaviour, reduction of reflexes, respiratory function, etc.).

The fish were considered to be dead when no vitality signs, such as movement of gills, escape reaction and body movement could be observed.

All animal studies were conducted in accordance with national law (D.lgs 26/2014) and European Directive 2010/63/UE on the protection of animals used for scientific purposes.

2.4 Data Analysis

Results of the toxicity tests on SDS are reported as Median-Lethal-Concentration (LC_{50}). LC_{50} values and their 95% confidence intervals were calculated by using Trimmed-Spearman-Karber (TSK) method version 1.5 (TOXStat software package) [39].

3. RESULTS

All the acute toxicity tests performed with sea bass exposed to SDS were considered valid, since no mortality was observed in all of the control groups, the range of pH values was always lower than 1,0 pH unit and the oxygen concentrations were always higher than 60 per cent of the air saturation value during the test.

Sodium Dodecil Sulfate nominal concentrations were used to calculate the median lethal concentrations at 96h (96h-LC₅₀), and their 95% confidence intervals, at two different salinities (20‰ and 5‰). The results are shown in the Table 3.

The analysis aiming at confirming the SDS test concentrations were not performed during our sea bass acute toxicity tests. However, we conducted the tests with a renewal of SDS solutions every 24 and 48 hours. Results obtained with a 24h renewal (96h-LC₅₀: 8.12 mg/l at salinity of 20‰ and 13.81 at salinity of 5‰) were comparable with those obtained with a 48h renewal (96h-LC₅₀: 9.50 mg/l at salinity of 20‰ and 14.14 at salinity of 5‰).

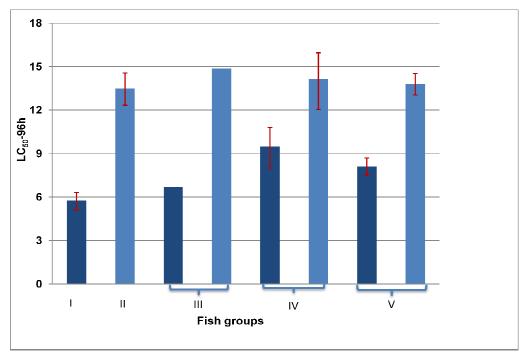


Fig. 1. Histogram of 96h-LC₅₀'s obtained with sodium dodecyl sulfate at two different salinities 20‰ (dark column) and 5‰ (light column) on five different sized sea bass (*Dicentrarchus labrax*) groups

Fish group	W	TL	96 h-LC₅₀ (95% Cl) mg/l					
	(g)	(cm)	20 ‰	5 ‰				
Ι	0.068±0.03	2.23±0.22	5.76 (5.20-6.40)	-				
II	0.50±0.07	3.90±0.30	-	13.50 (12.43 – 14.67)				
III	0.66±0.22	4.64±0.29	6.71 *	14.87 *				
IV	0.94±0.34	4.79±0.52	9.50 (8.2-11.0)	14.14 (12.32-16.24)				
V	1.42±0.05	5.40±0.07	8.12 (7.55-8.73)	13.81 (13.09-14.57)				

Table 3. Median Lethal Concentration (96h-LC₅₀) of sodium dodecyl sulfate for *Dicentrarchus labrax* juveniles at two different salinities (20 and 5‰)

Abbreviations: W: weight; TL: total length; CI: confidence interval;

Note: *The 95% CI is not reliable because no SDS concentration was found to prove an intermediate toxicity

As shown in Fig. 1, the SDS at salinity of 5‰ was always found to be significantly less toxic for juvenile sea bass independently of the fish size. In the present study, the reduction of salinity to 5‰ resulted in a decreased toxicity of SDS for all exposed fish groups (96h-LC₅₀: (II) 13.50 mg/l; (III) 14.87 mg/l; (IV) 14.14 mg/l; (V) 14.87 mg/l).

Significant differences were also seen (in Fig. 1) at a salinity of 20‰ among different fish groups. By raising the size of organisms from 2.23 ± 0.22 cm (fish group I) to 4.79 ± 0.52 cm (fish group V) the sensitivity of fish to SDS decreased (96h-LC₅₀: 5.76 mg/l vs 9.50 mg/l). On the other hand, no significant differences in toxicity to SDS were found between fish groups IV and V, probably due to similar size of sea bass.

4. DISCUSSION

The European sea bass was tested for chemical toxicity under many different conditions [40-47]. However, it is very important to understand how salinity might affect the toxicity of chemicals since euryhaline organisms can be employed in laboratory testing.

Some studies have demonstrated that sea bass is able to live in low salinity environments and to successfully adapt to freshwater under experimental conditions [48,49], even though in other studies mortality was recorded when fish were exposed to fresh water (<1‰) [50]. It was also found that sea bass juveniles have a low saline *preferendum* (15‰) which corresponds to the conditions this species encounter in the wild during their juvenile ecophase [51].

Moreover in order to examine the possible benefits of culturing sea bass at reduced salinities, Dendrinos and Thorpe [52] investigated the effects of various salinities (33, 30, 25, 20, 15, 10, 5‰) on growth and body composition in sea bass juveniles over a period of 1 year. They found out that all specimens were able to survive at reduced salinity percentages (until 5‰), whereas for those kept in freshwater, mortality occurred within the first 20 days. At salinities ranging from 18‰ to 8‰, *D. labrax* show lower locomotion and less agility and swimming activity in the tanks, in order to reduce food energy consumption and attribute larger portions for osmoregulation [53]. As salinity decreases (from 30‰ to 20, 10 and 5‰) the sea bass, in resting conditions, reduces its metabolic rates [54].

The toxicity of many chemicals was reported to vary according to water salinity. A critical review which investigated the effects of salinity on the toxicity of various classes of inorganic and organic chemicals to aquatic biota showed that negative correlations (such as toxicity increases while salinity decreases) are more frequent (55%) than positive (18%) and no correlations (27%) are also possible. The increasing toxicity with decreasing salinity observed with most metals (such as cadmium, chromium, copper, mercury, nickel and zinc) is probably due to the greater bioavailability of the free metal ion (toxic form) at lower salinity conditions [51]. Lower salinity (15‰) seems to increase the solubility of polycyclic aromatic hydrocarbons (PAH) from the oil dispersants, so that the exposure to PAH can be up to 60-fold greater compared to water of full salinity (32‰). On the other hand. organophosphate and carbamate insecticides toxicity appeared to decrease with decreasing salinity [55].

In the past, SDS has been often used by Italian laboratories as reference toxicant for assessing the sensitivity of *D. labrax* as well as for monitoring the condition of fish stocks and laboratory environments. The SDS toxicity data on sea bass from scientific literature, reported in Table 4, are referred at different life stages of sea bass, time exposures and salinities (from 20 to 35‰). In all reported studies, the LC₅₀ values are based on nominal SDS exposure concentrations.

	Sea bass				Test con	est conditions References References		References			
LS and age	W(g)	TL(cm)	T(C°)	Salinity (‰)	TP	Time exp	OECD TG	LC50 (mg/l)	±SD	95% CI (mg/l)	_
E 70-80 hpf	-	-	20±0.5	35	S	24h	203	< 1.12	-	-	Roncarati et al. [46]
L 30 d	0.005	0.011	20±0.5	20±1	S	24h	203	1.89	-	-	Roncarati et al. [46]
L 30 d	-	-	20±1	30±1	S	24h	-	2.04	-	1.38-2.52	Cicero et al. [63]
								2.90	-	2.64-3.18	
								2.66	-	1.99-3.55	
L 30 d	-	-	20±1	20±1	S	24h	203	1.89	-	1.65-2.15	Spaggiari et al. [23]
PL	0.06	-	20±0.5	15±1	S	24h	-	5.78	-	-	Roncarati et al. [46]
J	0.84	-	20±0.5	15±1	S	24h	-	9.85	-	-	Roncarati et al. [46]
J	3.40	-	20±0.5	15±1	S	24h	-	10.72	-	-	
J 50 d	-	1.21±0.14	21±1	30±1	S	24h	203	2.93	0.52	-	Mariani et al. [59]
						48h		0.90	0.50		
J 70 d	-	1.73±0.15	21±1	30±1	S	24h	203	3.98	0.99	-	Mariani et al. [26]
						48h		3.87	1.03	-	
J	-	5.75±0.2	20±1	20±1	R 48h	96h	203	7.28	0.47	-	Gelli et al. [56]
J	0.82±0.3	5.0±1.0	20±1	20±1	S	96h	203	7.07	-	5.0-10.0	Mariani et al. [57]
								7.88	-	7.10-8.19	
J	from 0.53±0.1 to 4.77±0.2	from 4.4±0.3 to 7.84±0.3	20±1	20±0.5	R 48h	96h	203	7.33	0.53	-	ICRAM-TAXAp- [24]

Table 4. Summary of Italian toxicity studies with sodium dodecyl sulfate on Dicentrarchus labrax at different life stages and test conditions

Abbreviations: CI: confidence interval; d: days; e-L: embryo-larvae; E: eggs; hpf.: hours post fertilization; J: juvenile; L: larvae; LS: life stage; P: Test Procedure; PL: post larvae; R: test with renewal of medium; S: static test; SD: standard deviation; TL: Total Length; W: weight. Note ^a Experiments were carried out by four laboratories with different batches of fish

Anderson et al. [61]

ICRAM Taxap-a [24]

Fish species	LS	S(‰)	Time exposure	LC₅₀ (mg/l)	SD	95% CI	References
Cyprinodon variegatus	L7d	32	7 d	2.9	-	1.8-3.4	Morrison et al. [65]
Menidia berillina	L 7 d	32	7 d	1.8	-	-	Morrison et al. [65]
Menidia berillina	L 28 d	20	96 h	1.48	-	1.35-1.63	Hemmer et al. [64]
Menidia menidia	-	10	96 h	2.8	-	2.55-2.98	Roberts et al. [60]
Cyprinodon variegatus	-	15	96 h	9.0	-	-	Anderson et al. [61]
Cyprinodon variegatus	-	10	96 h	4.1	-	3.83-4.47	Roberts et al. [60]
Gasterosteus aculeatus	-	20	96 h	0.51	-	-	McAuliffe CD, et al. [62]
				4.2	-	-	
Atheronops affinis	L 22 d	20	96 h	1.88	-	1.67-2.11	Hemmer et al. [64]
Fundulus heteroclitus	А	20	96 h	5.60	-	-	La Roche et al. [66]

Table 5. Summary of toxicity data with sodium dodecyl sulfate on different marine/estuarine fish species according to the IUCLID (2000) database of existing chemicals of the European Chemicals Bureau

Abbreviations: A: adult; d: days; h: hour; J: juvenile; L: larvae; LS: Life Stage; S: salinity; SD: Standard Deviation; CI: Confidence Interval. Note: ^aExperiments were carried out by four laboratories with 5 different batches of fish

15-20

20

-

J

96 h

96 h

Fundulus similis

Sparus aurata

4.70

4.50

7.66

0.81

The comparisons of results among different species should be considered with caution to avoid bias in the conclusions due to different conditions tested and possible confusing factors. Despite of a large quantity of SDS toxicity data on sea bass (Table 4), a stringent comparison with our data was not possible, because of so many different test conditions and life stages of fish were employed in the experiments. Our 96h-LC₅₀ values at salinity of 20‰ (9.50 and 8.12 mg/l) are comparable with those obtained from Gelli et al. [56], Mariani et al. [57] and ICRAM-TAXA, [24] with same sized sea bass juveniles. Furthermore, a positive correlation between toxicity and salinity might be suggested between the nominal 24h-LC₅₀ values at a salinity of 15‰ and those at a salinity of 30% [58] although the sizes of juveniles seem to be different.

It desirable that species be representative of the estuarine biota in sensitivity to toxic substances. In the present study, the reduction of salinity to 5‰ resulted in a decreased toxicity of SDS for all exposed fish groups. Moreover, these results confirmed our nominal 96h-LC₅₀ value of 13.4 mg/l (95% CI: 12.4-14.4 mg/l) obtained with Oncorhynchus mykiss exposed in similar test conditions (fish size: 3.16±1.04 g and 6.7±0.7 cm) (data not published) and nominal 96h-LC₅₀ values of 16.2 mg/l and 17.0 mg/l reported in the IUCLID database (2000) with Poecilia reticulata and Pimephales promelas. At a salinity of 5‰, D. labrax seems to show sensitivity to SDS which could be compared with freshwater fish species recommended by OECD TGs.

A summary of toxicity data with SDS on other marine fish species is reported in Table 5. Here again, we observed a great variability in the SDS LC_{50} values among species and also within species.

On the whole, the toxicity data derived from marine fish species are scarcely comparable with data on *D. labrax* because of different test, life stage and exposure conditions. The sea bass larvae with nominal 24h-LC₅₀ values ranging from 1.89 to 2.90 mg/l [63] seem to have a higher sensitivity in comparison with larvae of *Menidia* berillina, Cyprinodon variegatus [64] and Atherinops affinis [65] which show nominal 96h-LC₅₀ values of 1.8 mg/l, 2.9 mg/l and 1.88 mg/l, respectively. On the other hand, the nominal 96h-LC₅₀ value of 7.66±0.81 mg/L found in Sparus aurata (ICRAM Taxap-2005) [24] is

comparable with those of sea bass juveniles which ranging from 6.71 and 9.50 mg/L (our data) and from 7.07 to 7.88 mg/L [56,57,24].

Lastly, under similar conditions of salinity (20‰) and life stage, the sea bass seems to be less sensitivity in comparison with mummichog (*Fundulus heteroclitus*) which shows nominal 96h-LC₅₀ values of 5.60 and 4.70 mg/l [66].

5. CONCLUSIONS

In this study *D. labrax* confirms its ease of culture and maintenance under laboratory conditions, it is a well-known species, frequently used to investigate the toxicity of various classes of chemicals.

As euryhaline species, is able to adapting at different salinity condition both in nature and in laboratory conditions. In our experimental conditions juveniles sea bass at 5‰ salinity have sensibility of Rainbow trout comparable (O. mykiss) exposed to SDS in a similar test conditions. In this test conditions D. labrax seems to show sensitivity to SDS which could be compared with freshwater fish species recommended by OECD TGs. On the other hand, D. labrax shows an increasing sensibility at 20‰ water salinity; this means that if we change water salinity we get a more sensitive response from D. labrax. This characteristic could be helpful in case of use D. labrax like sentinel species for detecting environmental impact, like, for example, in assessing changing on water temperature due to climate change [67,68], on temperature increase due to water discharge from power plant [69], or studying the impact of toxic substances on the marine life like dispersant products (Gorbi et al. [1]. Tornambé et al. [14]) or priority substances (Roméo et al. [45]. Abreu et al., [19]. Antunes et al. [8]. Gravato et al. [16]. Banni et al. [10]. Ameur et al. [20]), as requested by Water Framework Directive.

The results of this study show that the characteristics of juveniles and adults of *D. labrax* make this species useful to provide information of toxicity in both seawater and freshwater. Its ease of maintenance and testing under laboratory conditions, and its broad euryhalinity confirm the convenience of use it as a sentinel species for detecting environmental impact and as a rule in monitoring studies.

6. FUNDING SOURCE

This research was funded by Italian Technical Committee of REACH (at Italian Ministry of Health) within the Project entitled "Inclusion of the European sea bass (*Dicentrarchus labrax* L. 1758) among species recommended for testing in the OECD Guidelines for testing of chemicals".

ACKNOWLEDGEMENTS

The authors would like to thank Pierluigi Trentini and Donatella Palazzi for organizing and performing activities at ARPA Ferrara.

The authors would like also to thank Ms Giulia Bernini for the English revision of this paper.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Parisi G, Terova G, Gasco L, Piccolo G, Roncarati A, Moretti VM, et al. Current status and future perspectives of Italian finfish aquaculture. Rev Fish Biol Fish. 2014;24:15-73. DOI:10.1007/s11160-013-9317-7.
- FAO. Cultured aquatic species information programme. FAO Fisheries and Aquaculture Department. Rome; 2013. Available:<u>http://www.fao.org/fishery/spec</u> <u>ies/2291/en</u> (Accessed on December 2013)
- Kennedy M, Fitzmaurice P. The biology of the bass, Dicentrarchus labrax, in Irish waters. J Mar Biol Assoc UK. 1972; 52:557–597.
- Vinagre C, Madeira D, Narciso L, Cabral H, Diniz M. Impact of climate change on coastal versus estuarine nursery areas: Cellular and whole-animal indicators in juvenile seabass Dicentrarchus labrax. Mar Ecol Prog Series. 2012;464:237–243.
- 5. Bœuf G, Payan P. How should salinity influence fish growth? Comp Biochem Physiol Part C. 2001;130:411-423.
- Sampaio LA, Wasielesky W, Miranda-Filho KC. Effect of salinity on acute toxicity of ammonia and nitrite to juvenile Mugil platanus. Bull Environ Contam Toxicol. 2002;68:668-674.
- 7. Ahmad I, Maria VL, Pacheco M, Santos MA. Juvenile sea bass (*Dicentrarchus*

labrax L.) enzymatic and non-enzymatic antioxidant responses following 17 bestradiol exposure. Ecotoxicology. 2009; 18:974-982.

- Antunes P, Hendriks AJ, Huijbregts MAJ, Gil O, Reis-Henriques MA. Organ-specific accumulation and elimination patterns of PCBs in adult seabass (*Dicentrarchus labrax*). Sci Total Environ. 2008;407:204-210.
- Bado-Nilles A, Quentel C, Mazurais D, Zambonino-Infante JL, Auffret M, Thomas-Guyon H, et al. *In vivo* effects of the soluble fraction of light cycle oil on immune functions in the European sea bass, *Dicentrarchus labrax* (Linné). Ecotoxicol Environ Safety. 2011;74:1896-1904.
- Banni M, Jebali J, Guerbej H, Dondero F, Boussetta H, Viarengo A. Mixture toxicity assessment of nickel and chlorpyrifos in the sea bass *Dicentrarchus labrax* L. Arch Environ Contam Toxicol. 2011;60:124-131.
- 11. Cotou E, Henry M, Zeri C, Rigos G, Torreblanca A, Catsiki V-A. Short-term exposure of the European sea bass *Dicentrarchus labrax* to copper-based antifouling treated nets: Copper bioavailability and biomarkers responses. Chemosphere. 2012;89:1091-1097.
- Danion M, Deschamps MH, Thomas-Guyon H, Bado-Nilles A, Le Floch S, Quentel C, et al. Effect of an experimental oil spill on vertebral bone tissue quality in European sea bass (*Dicentrarchus labrax* L.). Ecotoxicol Environ Safety. 2011; 74:1888-1895.
- De Domenico E, Mauceri A, Giordano D, Maisano M, Gioffrè G, Natalotto A, et al. Effects of "In vivo" exposure to toxic sediments on juveniles of sea bass (*Dicentrarchus labrax*). Aquat Toxicol. 2011;105:688-697.
- Della Torre C, Tornambè A, Cappello S, Mariottini M, Perra G, Giuliani S, et al. Modulation of CYP1A and genotoxic effects in European sea bass (*Dicentrarchus labrax*) exposed to weathered oil: A mesocosm study. Mar Environ Res. 2012;76,48-55.
- Gorbi S, Benedetti M, Virno Lamberti C, Pisanelli B, Moltedo G, Regoli F. Biological effects of diethylene glycol (DEG) and produced waters (PWs) released from offshore activieties: A multi-biomarker approach with the sea bass *Dicentrarchus labrax.* Environ Pollut. 2009;157:3166-3173.

- Gravato C, Guilhermino L. Effects of benzo(a)pyrene on seabass (*Dicentrarchus labrax* L.): biomarkers, growth and behaviour. Hum Ecol Risk Assess: An International Journal. 2009;15: 121-137.
- 17. Gravato C, Santos MA. *Dicentrarchus labrax* biotransformation and genotoxicity responses after exposure to a secondary treated industrial/urban effluent. Ecotoxicol Environ Safety. 2003;55:300-308.
- Hernandez-Moreno D, Pérez-Lopez M, Soler F, Gravato C, Guilhermino L. Effects of carbofuran on the sea bass (*Dicentrarchus labrax* L.): Study of biomarkers and behaviour alterations. Ecotoxicol Environ Safety. 2011;74:1905-1912.
- Abreu SN, Pereira E, Vale A, Duarte AC. Accumulation of mercury in sea bass from a contaminated Lagoon (Ria de Aveiro, Portugal). Mar Pollut Bull. 2000;40:293-297.
- Ameur WB, Trabelsi S, El Megdiche Y, Hassine SB, Barhoumi B, Hammami B, et al. Concentration of polychlorinated biphenyls and organochlorine pesticides in mullet (*Mugil cephalus*) and sea bass (*Dicentrarchus labrax*) from Bizerte Lagoon (Northern Tunisia). Chemosphere. 2013; 90:2372-2380.
- 21. Caruso G, Genovese L, Maricchiolo G, Modica A. Haematological, biochemical and immunological parameters as stress indicators in *Dicentrarchus labrax* and *Sparus aurata* farmed in off-shore cages. Aquacult Int. 2005;13:67-73.
- 22. Tornambè A, Giuliani S, Della Torre C, Amato E, Buffagni M, Catalano B, et al. Approccio ecotossicologico per la valutazione degli effetti su organismi marini dovuti ad inquinamento da idrocarburi in mesocosmo. Biol Mar Med. 2009;14(2):187-199. (Italian).
- Spaggiari R, Gelli F, Palazzi D, Pregnolato L, Venturini F, Savorelli F, et al. Sostanze prioritarie: I pesci (*Dicentrarchus labrax, Cyprinus carpio*) quali organismi bersaglio in test eco tossicologici, di bioconcentrazione e in saggi finalizzati a valutazioni di genotossicità. ARPA Emilia Romagna – Sezione Provinciale di Ferrara (Eds), Progetto Nazionale di monitoraggio acque superficiali; 2005. (Italian).
- 24. ICRAM TAXA. Programma di ricerca TAXA. Sperimentazione di test

tossicologici su organismi marini, ai fini dell'applicabilità del D.D. 23.12.2002. ICRAM Relazione finale Aprile 2005; 2005. (Italian).

- OECD. Fish, acute toxicity test Series on Guideline for testing of chemicals n° 203; 1992.
- Mariani L, Savorelli F, De Luca Picione F, Del Prete B, Di Capua E, Di Lorenzo B, et al. Standardization of acute toxicity biomassa protocol using European sea bass (*Dicentrarchus labrax*, L.1758): 2nd inter-laboratory comparison. Biol Mar Med. 2009;14:2:24-30.
- Gelli F, Cicero AM, Melotti P, Roncarati A, Pregnolato L, Savorelli F, et al. A proposal of method to evaluate the quality of marine waters: Optimization of 7 days bioassays using *Dicentrarchus labrax* (L.) juveniles. Chem Ecol. 2004;20:225-229.
- Manfra L, Maggi C, Bianchi J, Mannozzi M, Faraponova O, Mariani L, Onorati F, et al. Toxicity evaluation of produced formation waters after filtration treatment. Nat Science. 2010;2:33-40.
- 29. Mariani L, Manfra L, Maggi C, Savorelli F, Di Mento R, Cicero AM. Produced Formation Waters: A preliminary study on chemical characterization and acute toxicity by using fish larvae (*Dicentrarchus labrax*, L. 1758). Fres Environ Bull. 2004;13:1427-1432.
- Palazzi D, Altavilla E, Ferioli A, Gelli F, Savorelli F, Trentini PL. An ecotoxicological battery tests for the characterization of sediments to be dredged in Ravenna harbor. Biol Mar Med. 2009;14:52-56.
- OECD. Fish toxicity testing framework Series on Testing and Assessment n°171. ENV/JM/MONO. 2012;16.
- OECD. Fish, prolonged toxicity test: 14day study - Series on Guideline for testing of chemicals n°204; 1984.
- OECD. Fish, Juvenile Growth Test Series on Guideline for testing of chemicals n° 215; 2000.
- Ying GG. Fate, behavior and effects of surfactants and their degradation products in the environment. Environ Int. 2006;32:417-431.
- 35. Heerklotz H. Interactions of surfactants with lipid membranes. Q Rev Biophys. 2008;41(3-4):205-264.
- Schirmer K, Tanneberg K, Kramer NI, Völker D, Scholz S, Hafner C, et al. Developing a list of reference chemicals for

testing alternatives to whole fish toxicity tests. Aquat Toxicol. 2008;90:128-137.

- Steber J, Gode P, Gyhl W. Fatty alcohol sulfates. Soap Cosmet. Chem. Spec. 1988;64:44-50.
- George AL. Seasonal factors affecting surfactant biodegradation in Antarctic coastal waters: Comparison of a polluted and pristine site. Mar Environ Res. 2002;53:403-415.
- Hamilton M, Russo R, Thurston R. Trimmed spearman-karber method for estimating median lethal concentrations in toxicity bioassays. U.S. Environmental Protection Agency, Washington, D.C., EPA/600/J-77/178 (NTIS PB81191918); 1977.
- Basaran F, Sen H, Karabulut S. Effects of 2-phenoxyethanol on survival of normal juveniles and malformed juveniles having lordosis or nonfunctional swimbladders of European sea bass (*Dicentrarchus labrax*). Aquacult Res. 2007;38:933-939.
- Conti D, Martone C, Cadoni F, Bellaria V, Balzamo S, Belli M. Saggio di tossicità cronica sul branzino (*Dicentrarchus labrax* L.): influenza della stabilità chimica delle sostanze tossiche. Biologi Italiani. 2007;4:45-50. (Italian).
- 42. Diniz MS, Pereira R, Freitas AC, Rocha-Santos TAP, Castro L, Peres I, Duarte AC. Evalutation of the sub-lethal toxicity of belached kraft pulp mill effluent to *Carassius auratus* and *Dicentrarchus labrax.* Water Air Soil Pollut. 2011;217:35-45.
- 43. EI-Sayed YS, Khalil HR, Saad TT. Acute toxicity of ochratoxin-A in marine waterreared sea bass (*Dicentrarchus labrax* L.). Chemosphere. 2009;75:878-882.
- 44. Lemarié G, Dosdat A, Covès D, Dutto G, Gasset E, Person-Le Ruyet J. Effect of chronic ammonia exposure on growth of European seabass (*Dicentrarchus labrax*) juveniles. Aquaculture. 2004;229:479-491.
- 45. Roméo M, Bennani N, Ganssia-Barelli M, Lafaurie M, Girard JP. Cadmium and copper display different responses towards oxidative stress in the kidney of the sea bass *Dicentrarchus labrax*. Aquat Toxicol. 2000;48:185-194.
- 46. Roncarati A, Gelli F, Melotti P, Palazzi D, Pregnolato L, Savorelli F, et al. Impiego del branzino (*Dicentrarchus labrax* L.) nello svolgimento di test ecotossicologici: Effetti di un tossico di riferimento (sodio

laurilsolfato) su diversi stadi vitali. Riv Ital Acquacolt. 2001;36:43–52. (Italian).

- 47. Tornambè A, Manfra L, Mariani L, Faraponova O, Onorati F, Savorelli F, et al. Toxicity evaluation of diethylene glycol and its combined effects with produced waters of off-shore gas platforms in the Adriatic Sea (Italy): Bioassays with marine/ estuarine species. Mar Environ Res. 2012;77:141-149.
- Cataudella S, Allegrucci G, Bronzi P, Cataldi E, Cioni E, Corti M, et al. Multidisciplinary approach to the optimisation of sea bass (*Dicentrarchus labrax*) rearing in freshwater – Basic morpho-physiology and osmoregulation. European Aquaculture Society Spec. Publ. 1991;14:56-57.
- 49. Venturini G, Cataldi E, Marino G, Pucci P, Garibaldi I, Bronzi P, Cataudella S. Serum ions concentration and ATPase activity in gills, kidney and oesophagus of European sea bass (*Dicentrarchus labrax*, Pisces, Perciformes) during acclimation trials to fresh water. Comp Biochem Physiol A. 1992;103:451-454.
- Jensen MK, Madsen SS, Kristiansen K. Osmoregulation and salinity effects on the expression and activity of Na+/K+-ATPase in the gills of European sea bass, *Dicentrarchus labrax* (L.). J Exp Zool. 1998;282:290-300.
- Saillant E, Fostier A, Haffray P, Menu B, Chatain B. Saline preferendum for the European sea bass, *Dicentrarchus labrax*, larvae and juveniles: Effect of salinity on early development and sex determination. J Exp Mar Biol Ecol. 2003;287:103-117.
- 52. Dendrinos P, Thorpe JP. Effects of reduced salinity on growth and body composition in the European bass *Dicentrarchus labrax* (L.). Aquaculture. 1985;49:333-358.
- 53. Conides AJ, Glamuzina B. Laboratory simulation of the effects of environmental salinity on acclimation, feeding and growth of wild-caught juveniles of European sea bass *Dicentrarchus labrax* and gilthead sea bream, *Sparus aurata*. Aquaculture. 2006;256:235-245.
- 54. Claireaux G, Lagardère JP. Influence of temperature, oxygen and salinity on the metabolism of the European sea bass. J Sea Res. 1999;42:157-168.
- 55. Hall LW, Anderson RD. The influence of salinity on the toxicity of various classes of

chemicals to aquatic biota. Crit Rev Toxicol. 1995;25:281-346.

- 56. Gelli F, Savorelli F, Floris B, Roncarati A, Pregnolato L, Trentini PL, et al. Use of the sea bass (*Dicentrarchus labrax*, L.) as biological target in toxicity and bioconcentration tests. Biol Mar Med. 2005;12:684-687.
- 57. Mariani L, De Pascale D, Faraponova O, Tornambè A, Sarni A, Giuliani S, et al. The use of a test battery in marine ecotoxicology: The acute toxicity of sodium dodecyl sulfate. Environ Toxicol. 2006; 21:373-379.
- 58. Marques E, Miguel M, Dias R, Melnikov S, Khan A, Lindman B. Gel formation and association in systems of cationic surfactant vescicles and oppositely charged polymers. Polymer Preprints 2000;41:737-738.
- 59. Mariani L, Savorelli F, Bellaria V, Cadoni F, Cigar M, De Luca Picione F, et al. Use of the European sea bass (*Dicentrarchus labrax*, L. 1758) in comparison exercises sperimentation for the validation and normation of the method for acute toxicological tests using Mediterranean fish species. Biol Mar Med. 2007;14(1):73-77.
- Roberts MH Jr., Warinner, JE, Tsai C-F, Wright D, Cronin, LE. Comparison of estuarine species sensitivities to three toxicants. Arch Environ Contam Toxicol 1982;11:681-692.
- Anderson JW, Neff JM, Cox BA, Tatem HE, Hightower GM. Characteristics of dispersions and water-soluble extracts of crude and refined oils and their toxicity to estuarine crustaceans and fish. Mar Biol 1974;27:75-88.
- 62. McAuliffe CD, Smalley AE, Groover RD, Welsh WM, Pickle WS, Jones GE. Chevron main pass block 41 oil spill: chemical and biological investigations. In: Proceedings Conference on prevention

and control of oil pollution. San Francisco, California. American Petroleum Institute, Washington D.C. 1975;555-566.

- 63. Cicero AM, Mariani L, Savorelli F, Roncarati A, Gelli F, Palazzi D, Pregnolato L. Preliminary trials for determining intralaboratory variability in ecotoxicological bioassays using *Dicentrarchus labrax*. Biol Mar Med. 2004;11:496-498.
- 64. Hemmer MJ, Middaugh DP, Comparetta V. Comparative acute sensitivity of larval topsmelt, *Atherinops affinis*, and inland silverside, *Menidia beryllina*, to 11 chemicals. Environ Toxicol Chem. 1992; 11:401-408.
- Morrison G, Torello E, Comeleo R, Walsh R, Kuhn A, Burgess R, et al. Intralaboratory precision of saltwater shortterm chronic toxicity tests. Res J Water Poll Contr Fed. 1989;61:1707-1710.
- LaRoche G, Eisler R, Tarzwell CM. Bioassay Procedures for Oil and Oil Dispersant Toxicity Evaluation: Journal Water Pollution Control Federation 1970;42:1982-1989.
- Person-Le Ruyet J, Mahé K, Le Bayon N, Le Delliou H. Effects of temperature on growth and metabolism in a Mediterranean population of European sea bass, *Dicentrarchus labrax*. Aquaculture. 2004; 237(1-4):269-280 Available:http://dx.doi.org/10.1016/j.aquac

68. Pope EC, Ellis RP, Scolamacchia M, Scolding JWS, Keay A, Chingombe P, Shields RJ, Wilcox R, Speirs DC, Wilson RW, Lewis C, Flynn KJ. European sea bass, *Dicentrarchus labrax*, in a changing

ocean. Biogeosciences. 2014;11,2519-

2530
69. Pawson MG, Eaton DR. The influence of a power station on the survival of juvenile sea bass in an estuarine nursery area. J. of Fish Biol. 1999;54(6):1143–1160.

© 2015 Conti et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://sciencedomain.org/review-history/11775