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Cactaceaes of the Brazilian Semiarid: Source of Bioactive Compounds

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Authors' contributions

This work was carried out in collaboration between both authors. Author LMRA carried out the material preparation, structural arrangement and wrote the manuscript. Author LGF coordinated the development of the study, and contributed to all parts and critical reading of the manuscript. Both authors read and approved the final manuscript.

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Review Article

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ABSTRACT

The semiarid region of Brazil stands out as one of the spaces with the highest diversity of plant species in the world, and the Cactaceae family the one that best represents the Brazilian semiarid. However, although there are many species of cacti in Brazil, their chemical potential has yet to be discovered. Given this, the present review aims to record the bioactive metabolites of native cactaceaes or not in Brazil, encompassing a description of its habitat and traditional uses. Compilations of ethnobotanical studies point to the importance of cacti species in the daily life of local cultures. Cactaceaes are used for food, economic, ornamental, and mystical purposes, among others, and stand out for their importance in traditional medicine, used to treat various diseases. Bioactive compounds in this family belong mainly to alkaloid groups, such as betalain, phenolic acids, terpenes and fatty acids. This review displays the relevance of the Cactaceae family in the face of the remarkable production of bioactive compounds.

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1. INTRODUCTION

Cactaceae is the term in the Latin language attributed to the family of plants that belong to the cacti. Cactaceaes are native to the American continent, having their most extensive st distribution in the arid and semiarid regions of the Americas. This family has approximately 1.500 species and 130 genera, divided into four subfamilies, of which three, Pereskioideae, Opuntioideae, and Cactoideae, occur in Brazil [1]. In the Brazilian territory, 37 genera and 275 species are recognized so far, of which 14 genera and 188 are endemic [2].

The geographical space of the semiarid, where it has the largest distribution of cactaceaes in Brazil, extends to eight states in the Northeast Region (Alagoas, Bahia, Ceará, Paraíba, Pernambuco, Piauí, Rio Grande do Norte and Sergipe) plus the north of Minas Gerais [1]. In this area, the dominant biome is caatinga, characterized by low precipitation, high temperature, and xerophyte vegetation [3].

Anatomical, ecological, and physiological aspects peculiar to the Cactaceae family are responsible for adapting it to the semiarid climate. Its efficiency in water use is based on the Specialized Photosynthetic mechanism CAM (Crassulacean Accident Metabolism), where the stomats open at night, with the lower temperatures, and keep them closed during the day, enabling high efficiency of the use of water [4].

Cactus species are usually xerophytic, succulent and perennial. They have arboreal, shrubby, subshrub, climbing, epiphyte, or geophyte strata and fibrous or tuberous roots [5]. They have a photosynthetically active stem of variable color, shape and size, forming cladodes, which can be smooth, cylindrical, columnar or globular and are usually covered with thorns [6].

The various cactaceae found in the Brazilian semiarid are used and known for various purposes, such as food, medicine, forage, religious mystical, for ornamentation, biodiversity conservation and as a rain bioindicator [7]. In the scope of traditional medicine, *Cereus jamacaru* (mandacaru), *Opuntia ficus-indica* (palm), *Melocactus zehntneri* (friar's crown), *Pilosocereus gounellei* (xique xique), *Pilosocereus pachycladus* (facheiro) and *Opuntia*

palmadora (palmadora), used by rural communities for back problems, diabetes, rheumatism, urinary tract infections, kidney problems, appendicitis, bronchitis and flu [8,9]. The traditionally used parts are mainly the cladodes and roots, in the form of infusion and decoction [10].

In consonance with its applicability in traditional medicine, these plants have important metabolic characteristics, such as the production of bioactive compounds. The accumulation of these compounds is one of the response mechanisms to biotic agents and abiotic stresses, which, as stimuli, trigger the synthesis of alkaloids, terpenes, betalains, phenolics and nitrogenous compounds [11].

However, although there are many species of cacti in the Brazilian semiarid region, their phytochemical potential still needs to be discovered. Ttherefore, be the subject of study given these plants' importance. In this sense, the present review aims to identify the bioactive metabolites of cacti, whether or not native to Brazil, encompassing a description of their habitat and traditional uses. To this end, a search was carried out in scientific databases, where important publications concerning cacti were extracted.

2. CACTACEAE FAMILY - HABITAT AND TRADITIONAL USES

The Cactaceae family belongs to the Angiosperms group, divided into three subfamilies: Opuntioideae, Pereskioideae and Cactoideae – the latter is the most numerous in genera [12]. However, a fourth subfamily, Maihuenioideae, has also been recognized by some authors, being restricted to South America (Argentina and Chile) [13]. This family is native to the American continent, where it occurs with high richness and abundance of species, from British Columbia and Alberta in Canada to Patagonia in Argentina, including the island regions of the continent. Few cactus species are seen occurring naturally on other continents [14].

Four areas in the world are identified as having high richness and endemism of species; they are: 1) Mexico and the southwest of the USA; 2) the central Andes region involving Peru, Bolivia, southern Ecuador, northeastern Chile, and northwestern Argentina; 3) Eastern Brazil,

covering the northeastern region and part of the southeastern region, excluding the southern portion of the State of Rio de Janeiro and the entire State of São Paulo; and 4) the region that includes Paraguay, Uruguay, northern Argentina and Brazil [1].

The term cactus is used to designate the group of species belonging to the Cactaceae family, which have morphological and physiological adaptations that collaborate with the conservation of species diversity in northeastern Brazil. Cacti generally have a thick cuticle, mucilaginous tissues, succulent and thorny stems, leafless branches and areolas producing thorns [3,15].

As these plants develop in adverse
environments. accumulating secondary accumulating secondary metabolites are one of the response mechanisms to environmental stress. Thus, the water deficit, the high saline concentration in the soil and the light stress are stimuli that trigger the synthesis of compounds such as terpenes, phenolics and nitrogen compounds [15].

The Caatinga species of Cactaceae are classified as xerophilous; they have specific characteristics for semiarid regions, such as the

ability to tolerate water scarcity and resist the drought period [3]. Therefore, these plants have a specialized survival mechanism, with physiological adaptations to high temperatures. These adaptations are related to the ability of cacti to convert water into dry matter through a specialized photosynthetic mechanism called MAC (Crassulacean Acid Metabolism), which allows them to remain succulent during the dry season [16,17].

Crassulacean Acid Metabolism (CAM) is one of three possible atmospheric carbon (CO_2) assimilation types via photosynthesis. The CAM plants have increased water use efficiency, constituting a necessary physiological adaptation that allows the plants to occupy habitats characterized by intermittent water availability [16,17].

The Cactaceae is among the most used plants by man, mainly in the semiarid region. The diversity of uses brings it into four categories: medicinal, food, ornamentation, fuel, and fodder plant (Table 1). In this way, one can notice that this botanical group has excellent economic, biological and ecological value, mainly because it always remains green, even in water stress conditions and under high temperatures [18].

Table 1. Traditional uses of cactaceae from the Brazilian semiarid region

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The robust structure of several cacti, such as *C. jamacaru* DC. and *Echinopsis attackamensis* (Phil.), makes them suitable for building fences, battens, boards, doors and windows [19,9]. Other uses, such as the mucilage of the cactus *Opuntia ficus* and *Pereskia aculeata*, have been applied in the food packaging industry as a raw material for films and coatings. It has been used, more recently, as a food preservative [20].

One of the prominent uses for Cactaceae species is in folk medicine, food and forage [21,22]. According to Andrade *et al.* (2006b) [21], traditional medicine using cactus is ancient in the semiarid region and is based on traditional knowledge passed down through generations. In the same study, the author mentions the use of cactus for various health problems recorded by the research participants, namely: "heat", vaginal inflammation, urinary infection, flu, inflammation in the uterus, bellyache, "choking", "dryness", back pain, syphilis, kidney problem, urethra problem, colic, bowel problem, prosthesis problem, toothache, "swollen belly" and dysentery".

In human food, the ingestion of fruits can be mentioned, mainly of the genera *Cereus* and *Pilosocereus*. Their consumption can be for generating some products or in nature [23,24]. Given that, Mizrahi (2014) [23] conducted a study with the fruits of *Cereus peruvianus*, using the ripe fruit to develop jam, dried fruit and aromatic liqueur.

Furthermore, according to Shetty *et al.* [22], cacti have excellent properties; they have flavour, are nutritious and can be eaten fresh, as vegetables and in salad dishes. Food products such as cookies, candies, puddings and cakes have also been developed using the cactus. In addition, other commercial products can be mentioned, such as shampoos and soaps, produced from different species of this family [9,18]. Cactaceae is also used in animal feed, such as cattle and goats. The high content of vitamin A and iron present in these plants can meet the nutritional needs the region's animals, especially in drought [8].

However, due to the popularity of Cactaceae as a medicinal plant, studies have been developed to evaluate the chemical composition of these plants. Since then, alkaloids, carbohydrate polymers, phenolic compounds, carotenoids, natural pigments and terpenes have been described in Cactaceae, especially those belonging to the subfamilies of Cactoideae [18,25].

Considering the great diversity of use and the variety of cactus species, studies that focus on the biochemical characterization of this family are of paramount importance since factors such as habitat destruction threaten these species, putting them in danger of extinction [26]. The Red List of Threatened Species of the International Union for the Conservation of Nature (IUCN) includes about 139 cactus species as vulnerable to extinction [27],

highlighting the subfamilies Cactoideae and Opuntioideae with the most significant number of species represented on the IUCN Red List. Brazil has the highest proportion of threatened species on a global scale with 18%, followed by Mexico (10%), Ecuador (9%) and Peru (7%) [14].

3. BIOACTIVE COMPOUNDS OF CACTACEAE

Cacti contain many chemical substances with pharmacological and biological relevance, synthesized in response to biotic and abiotic factors. The main compounds produced are alkaloids [30], betalains [31] and phenolic compounds [32], terpenes [33] (Table 2) and fatty acids [34].

3.1 Alkaloids

Alkaloids are a heterogeneous group of nitrogenous substances, of fundamental character and pharmacological action, generally of plant origin. Approximately 12.000 compounds are present in plants and act as the defense mechanism against herbivores and some species of pathogens. Notably, the concentration of alkaloids in plants can be highly variable. Several factors, such as genetics, environment, age, climate, time of year or time of day, growth stage and wild or cultivated plants are considered capable of influencing the content and/or composition of alkaloids in plants [15].

Several alkaloids with important biological properties have already been identified in cacti. Among them is mescaline, present mainly in *Lophophora williamsii*, *Lophophora diffusa*, *Trichocereus pachanoi*, *Trichocereus peruvianus* and *Trichocereus bridgesii*; pellotine, present in *Lophophora diffusa*, and hordenine, present in the genera *Turbinicarpus*, *Mammillaria* and *Ariocarpus*. These alkaloids have been widely investigated due to their hallucinogenic properties, among other biological effects of interest [28]. However, these species and genera are not native to Brazil, and are not cultivated in the country [1].

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Other alkaloids have been identified in cacti, about 50 phenethylamines and about 80 isoquinolines, N-methyltyramine, tyramine and macromerine, which are the most commonly found in these plants [28]. In the genus Cereus, common in the Brazilian semiarid region, the most abundant alkaloids are N-methyltyramine (an adrenergic agonist), hordenine (stimulating norepinephrine) and tyramine (precursor of dopamine) (Fig. 1) [15]. N-Methyltyramine has also been identified in *Turbinicarpus alonsoi*, *Obregonia denegri* and *Lophophora williamsii*. This compound is related to its beneficial effect on treating gastrointestinal disorders [34,35].

In the species Cereus jamacaru, endemic to Brazil, with greater geographic distribution in the northeast region, hordenine, tyramine, Nmethyltyramine, Tyrosine and phenethylamine have already been reported [38].

3.2 Betalains

Betalains are nitrogenous and water-soluble substances that belong to alkaloids consisting of betacyanins and betaxanthins and are classified as vacuolar pigments. These compounds have many applications in the food, cosmetics and pharmaceutical industries, and can be used as a natural dye, constituting an alternative to synthetic dyes [44,45].

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Fig. 1. Chemical structures of the main nitrogen-containing compounds identified in the genera *Cereus* **and** *Opuntia***. (Images available at PubChem)**

These compounds are mainly produced by the genus Opuntia, and by other genera such as *Hylocereus, Mammillaria and Schlumbergera* [31,46]*.* The betalain profile of *O. ficus-indica var sanguigna* was described by Melgar et al. (2017)

[40] using the LC-DAD-ESI/MS technique. The authors identified seven betalain fractions, two betaxanthins (indicaxanthin isomer I, indicaxanthin isomer) and five betacyanins (Betanidin-5-O-β-sophoroside, Betanidin-5-O-β glucoside (betanin), Isobetanin, Gomphrenin I and Betanidin) (Fig. 1).

Betalains were also identified in fruits of the species *Stenocereus pruinosus* and *S. stellatus* by HPLC-DAD ESI/MS, revealing the predominance of indicaxanthin, gomphrenin I, phyllocacthin and their isomers, with betaxanthin content being higher than that of betacyanins in both species [46].

Studies demonstrate that cactus fruits have a more significant predominance of betalains thanclododium and seeds. For example, the fruit of *Hylocereus polyrhizus*, specifically in the fruit peel, were identified as betanin, isobetanin, phyllocactin, butyrylbetanin, hylocerenin, isophyllocactin, isobutyrylbetanin, 20-apiosylphyllocactin and 20-apiosyl isophyllocactin [47]. In fruits of *Mammillaria spp.,* the presence of betacyanins has been described, such as: betanidin 5-O-β-sophoroside, isobetanidin 5-O-βsophoroside, betanin, isobetanin, betanidin 5-O- (60-O-malonyl)-β sophoroside, isobetanidin 5-O- (60-O-malonyl)-β-sophoroside, betanidin 5-O- (40-Omalonyl)-β-sophoroside, isobetanidin 5-O- (40 Omalonyl)-β-sophoroside, phyllocactin, isophyllocactin, 40-O-malonyl-betanin, 40-Omalonylisobetanin, 20-O-apiosyl-phyllocactin and 20-Oapiosyl-isophyllocactin [48].

3.3 Phenolic compounds

The main phenolic compounds described in the cactus are phenolic acids, flavonoids, tannins, coumarins, lignans and stilbenes. These compounds have essential biological functions related to their chemical structure, consisting of a benzene ring with one or more hydroxyl groups [49,31,50].

A wide variety of phenolic compounds has already been described in the genus *Opuntia*, such as catechin [31,51], chlorogenic acid [31,51], ferulic acid [31], p-coumaric acid, quercetin, and its derivatives [52], kaempferol, isorhamnetin, gallic acid [54,53], protocatechuic acid, salicylic acid, rutin [52], kaempferol-3-O-rutinoside [53] (Fig. 2). *Opuntia ficus-indica* is one of the most studied species due to its metabolite profile; phenolic compounds are present in the cladode, fruits, and flowers, both in wild and cultivated species [54].

Mena et al. [50] analyzed the presence of
phenolic compounds. through liquid compounds, through liquid chromatography-mass spectrometry (UHPLC-ESI-MS) in the clododium and fruit of *Opuntia ficus-indica* (L.) Mill*.* of different cultivars. The results showed that the highest content of phenolic compounds was identified in young cladodes, with 26 compounds, with flavonoids (in particular, flavonols) being the leading group of polyphenolics. Identifying isorhamnetin, rutin, and ferulic acid-hexoside derivatives was also possible. Regarding the peel and pulp of the fruits, 26 phenolic compounds were identified in the peel, with a predominance of phenolic acids in detriment of flavonols; in the pulp, the amount was smaller, with 21 phenolic compounds, with a ferulic acid derivative being the major representative. In the fruit peel of most cultivars, the following compounds were identified in high concentrations: ferulic acid hexoside, sinapic acid-hexoside, dihydrosinapic acid hexoside and isorhamnetin-rutinoside.

The fruits of *Pilosocereus arrabidae* were also evaluated for the composition of phenolic compounds. Gonçalves *et al.* [49] evaluated the peel and pulp of *Pilosocereus arrabidae* by HPLC-MS, identifying the presence of important flavonoids: Catechin, Dihydrokaempferol, Quercetin, Quercetin 3 or 4′-O-glucoside and Rutin, in both botanical parts (Fig. 2). The high rutin content was found in the pulp of *Stenocereus pruinosus* and *S. stellatus* fruits [48], as well as in the peel of fruits of *Hylocereus undatus* [54].

The main phenolic compounds present in cactus flowers are phenolic acids and flavonoids. Ammar *et al.* [55] evaluated the flowers of *Opuntia ficus-indica* using the LC-MS/MS technique, identifying quinic acid as the main phenolic acid, followed by gallic, protocatechuic, chlorogenic, 4-O-caffeoylquinic, caffeic, p coumaric, trans ferulic and rosmarinic acids. Among the flavonoids, they identified kaempferol-3-O-rutinoside, rutin, hyperoside, 4,5 di-O-caffeoyl quinic acid, quercetin-3- Orhamoside, isorhamnetin-3-O-rutinoside, isorhamnetin, 3-O-glucoside, apegenin and kaempferol 3-O-arabinoside. It is important to note that most studies have focused on the cladode and fruit, with few studies focused on flowers.

Catechin $(C_{15}H_{14}O_6)$

P-Coumaric acid $(C_9H_8O_3)$

Isorhamnetin $(C_{16}H_{12}O_7)$

Salicylic acid $(C_7H_6O_3)$

Chlorogenic acid $(C_{16}H_{18}O_9)$

Quercetin $(C_{15}H_{10}O_7)$

Gallic acid $(C_7H_6O_5)$

Rutin $(C_{27}H_{30}O_{16})$

Kaempferol-3-O-rutinoside $(C_{27}H_{30}O_{15})$

Protocatechuic acid (3,4-

Dihydroxybenzoic acid)

 $(C_7H_6O_4)$

Ferulic acid

 $(C_{10}H_{10}O_4)$

Kaempferol

 $(C_{15}H_{10}O_6)$

Dihydrokaempferol (Aromadendrin) $(C_{15}H_{12}O_6)$

Fig. 2. Chemical structures of the main polyphenols identified in the Cactaceae family, which occur in the Caatinga. (Images available at PubChem)

Despite the presence of polyphenols being reported mainly in *Opuntia* species, particularly in *O. ficus-indica*, in other genera such as *Hylocereus* [56], *Pereskia* [26] and *Coryphantha* [57], the presence of these compounds is also
described. Gallic, vanillic, syringic, described. Gallic, vanillic, syringic, protocatechuic, p-hydroxybenzoic, p-coumaric and caffeic acids have been described in the

species *Hylocereus undatus* and *Hylocereus polyrhizus* [46,56].

3.5 Terpenes

Terpenes can be defined as "natural alkenes"; they have a carbon-carbon double bond and are characterized as an unsaturated hydrocarbon. Despite having structural differences, all terpenes/terpenoids are structured in five-carbon blocks – isoprene units (C_5H_8) – usually linked together in the "head-to-tail" order (link 1-4) [15].

Many terpenes have already been identified in cacti - in the cladode, stem, peel, seed and fruit, especially in the genus *Opuntias*, mainly in the species *Opuntia comonduensis* [58,59]*, Opuntia dilleniid* [50]*, Opuntia ficus-indica* [31], *Opuntia humifusa* [58]*, Opuntia littoralis* [32]*, Opuntia macrorhiza* [60]*, Opuntia polyacanta* var. arenaria [61] and *Opuntia phaeacantha* [61]*.* However, the presence of terpenes has also been reported in other genera, such as *Echinopsis* [58], *Hertrichocereus* [62], *Isolatocereus* [62], *Machaerocereus* [63], *Hylocereus* [56], *Pereskia* [25] and *Trichocereus* [62].

In the cladode of *Opuntia ficus-indica*, the presence of α-Tocopherol, β-tocopherol, γtocopherol, linoleic acid, palmitic acid, lauric acid and myristic acid (Fig. 3) was identified with HPLC-UV [32,31].

Linoleic, oleic, palmitic acids, cholesterol, campesterol, stigmasterol, and β-sitosterol have been described in *Hylocereus* seeds [56]. In the whole plant of *Isolatocereus dumortieri*, the presence of Dumortierinoside A (sapogenin), dumortierinoside A methyl ester, pachanoside I1 and pachanoside D1 was reported [62,64]. In the genus *Echinopsis*, specifically in the stem, there has already been the identification of: pachanosides C1, E1, F1 and G1 (1–4), bridgesides A1, C1, C2, D1, D2, E1, E2, 24- Methyl-cholesterol and sitosterol [58]. Bridgesigenin A and bridgesigenin B have also been described in the stem of the genus *Trichocereus* [62].

One can notice a wide variety of terpenes in Cactaceae species, as a diversity of triterpenoids, including unusual sterols, pentacyclic triterpenoids and saponins. These compounds, such as the terpenes of *Pereskia grandiflora*, have been studied for their biological activity. Sri Nurestri *et al.* [65] demonstrated that terpenes from the leaves of this species had an *in vitro* cytotoxic effect against five human carcinoma cell lines: nasopharyngeal, cervical, colon (HCT 116), hormone-dependent breast (MCF7) and lung (A549).

Isolatocereus dumortieri and *Stenocereus alamosensis* were also evaluated, demonstrating that triterpene saponins were related to type I antiallergic activity, detected by the inhibitory activity of the β hexosaminidase release from RBL-2H3 cells [65].

3.6 Other Compounds

Besides the compounds mentioned above, other biocompounds such as saponins, carotenoids, carbohydrate polymers and unsaturated fatty acids were detected in cactus plants [64,25,66,33].

Cacti, particularly from the Cactioideae family, are rich in saponins. Kakuta *et al.* [64] isolated three triterpenoid saponins in *Isolatocereus dumortieri* Backbg, being dumortierinoside A methyl ester, pacanoside I1 and canoside D1, in addition to two more saponins isolated from *Stenocereus alamosensis* A. C. Gibson & K. E. Horak, being gummy side A and gummy methyl ester A. Other saponins such as stellatoside (sapogenin glycosides) have been identified in *Stenocereus stellatus* [67] and stellatoside B and erucasaponin A (betulinic acid glycoside) in other species of the genus *Stenocereus* [68].

The presence of carotenoids was reported mainly in the species *Pereskia aculeata* Mill and *Pereskia grandifolia* Haw, highlighting high levels of β-carotene and α-carotene in berries of *Pereskia aculeata*, and high levels of β-carotene and xanthophylls (lutein and xanthophylls) in leaves of both species [25]. *Pereskia bleo*, lutein and zeaxanthin were also identified in berries [69].

The Cactaceae family is especially rich in mucilage, with a high content of carbohydrates along with water-soluble O-glycosides (such as flavonols and saponins). Arabinogalactans-like polymers, pectin and oligosaccharides have already been described in cacti [70]. Martin *et al.* [70] evaluated the mucilage of the leaves of *Pereskia aculeata* using the GC-MS and NMR technique, revealing the presence of galactose, arabinose, rhamnose, fucose and partially esterified galacturonic acid, making it possible to characterize a type I arabinogalactan.

Fig. 3. Chemical structures of the main terpenes and fatty acids identified in the Cactaceae family, which occur in the Caatinga. (Images available at PubChem)

Studies have also reported the presence of fatty acids in cacti. Benattia *et al.* [33] analyzed the fatty acids in *Opuntia ficus-indica* seeds by GC- MS, revealing the presence of linoleic (C18:2) and oleic (C18:1) fatty acids. The presence of these fatty acids was also reported by Ciriminna et al. (2017) [66] in the seed oil of *Opuntia ficusindica*, who also identified the presence of Sterols, such as b-sitosterol, Campesterol and Stigmasterol, in addition to Vitamin E (gtocopherol).

4. CONCLUSION

There is much to be explored with the cacti of the Brazilian semiarid region, both with the cladodes and with the fruits and flowers. These plants have phytochemical potential for obtaining bioactive compounds of interest.

Among the cacti evaluated in this review, the genera Opuntia, Pereskia and Pilosocereus are the most studied and documented for their antioxidant capacity and biological properties. The presence of phenolic compounds, betalains and terpenes, saponins, carotenoids, carbohydrate polymers and unsaturated fatty acids, confers its effectiveness as a medicinal/functional and food plant.

The Opuntia ficus-indica species is one of the most investigated, demonstrating a promising profile of bioactive compounds, especially phenolics and Betalains. Cereus jamacaru, native to Brazil, also stands out in its a range of bioactive compounds, are related to antioxidant, antitumor and antimicrobial activities.

However, among the vast universe of cacti that occur in the caatinga biome, few species that have already been studied and documented in the literature. Many of these may also present functional properties.

Studies evaluating this family's functional potential of this family can undoubtedly contribute to adding knowledge and value to the species. Because they are mainly rich in phenolic compounds and betalains, cacti have recognized antioxidant action and are related to health promotion. Thus, further scientific investigations can contribute to characterizing the plant and consequently stimulating the production and consumption of cacti as food.

Finally, the present review highlighted the variety of uses of the cactus, provided a broad view of its habitat and explored the immense phytochemical potential of these plants. Particularly notable are the bioactive compounds reported here, however, more in-depth studies, in vivo, are still needed to prove their efficacy and safety of use.

CONSENT

It is not applicable

ETHICAL APPROVAL

It is not applicable

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Cavalcante A, Teles M, Machado M. Cacti from the Semi-Arid region of Brazil: illustrated guide. National Semiarid Institute – INSA; 2013.
- 2. Zappi D, Taylor NP. Diversity and endemism of Cactaceae in the Espinhaço Range. Megadiversity. 2008;4(1):111-116.
- 3. Judd WS, Campbell CS, Kellogg EA, Stevens PF, Donoghue MJ. Plant systematics: a phylogenetic approach (3th ed.). Artmed, Porto Alegre; 2009.
- 4. Duke JG. The Northeast and xerophilic crops. 4th edition. Fortaleza: Banco do Nordeste do Brasil; 2004.
- 5. Zappi D, Taylor NP. Cactaceae in Flora of Brazil. Rio de Janeiro Botanical Garden; 2020. Accessed 25 March 2023.

Available:https://floradobrasil2020.jbrj.gov. br/FB70

- 6. Barroso GM, EF Guimarães, Ichaso CLF, Costa CG, Peixoto AL. Systematics of angiosperms in Brazil. Rio de Janeiro: Technical and Scientific Books Ed; 1978.
- 7. Silva VA, Diversity of use of cacti in northeastern Brazil: A review. Gaia Scientia. 2015;9(2):175-182.
- 8. Andrade CTS, Marques JGW, Zappi DC. Use of cacti by Bahian country people. Connective types to define utility

categories. Sitientibus, Biological Sciences Series. 2006a;6:3-12.

- 9. Lucena CM, de Lucena RFP, Costa GM, Carvalho TKN, da Silva CGG, da Nóbrega ARR, et al. Use and knowledge of Cactaceae in Northeastern Brazil. J. ethnobiol. ethnomed. 2013;9(1):1-11. Available:https://doi.org/10.1186/1746- 4269-9-62
- 10. Lucena CM, Carvalho TK, Ribeiro JE, Quirino ZG, Casas A, Lucena RFP. Traditional botanical knowledge about cacti in semi-arid Brazil. Gaia scientia. 2015;9(2):77-90.
- 11. Bartwal A, Mall R, Lohani P, Guru SK, Arora S. Role of secondary metabolites and brassinosteroids in plant defense against environmental stresses. J. Plant Growth Regulation. 2013;32:216–232.
- 12. Britton NL, Rose JN. The Cactaceae: descriptions and illustrations of plants of the cactus Family, Courier Corporation. 1963;3:263.
- 13. Souza VC, Lorenzi H. Systematic botany: Illustrated guide for identifying Angiosperm families in the Brazilian flora, based on APG II. 2nd edition. Nova Odessa: Instituto Plantarum; 2005.
- 14. Ortega-Baes P, Sühring S, Sajama J, Sotola E, Alonso-Pedano M, Bravo S, Godínez-Alvarez H. Diversity and conservation in the cactus family. Desert plants. 2010;157-173. Available:https://doi.org/10.1007/978-3- 642-02550-1_8
- 15. Trout K. Cactus chemistry by species (1th ed.). Mydriatic Productions; 2014.
- 16. Germano RH, Barbosa HP, Costa RG, Medeiros AN, Carvalho FFR. Assessment of the chemical and mineral composition of cacti in the semi-arid region of Paraíba. Technical agriculture. 1999;20(1):51-57.
- 17. Millar HA, Siedow JN, Day D. Respiration and photorespiration. In: Buchanan BB, Gruissem W, Jones RL, editors. Biochemistry and Molecular Biology of Plants. Berkeley: John Wiley & Sons; 2015.
- 18. Das G, Lim KJ, Tantengco OAG, Carag HM, Goncalves S, Romano A., et al. Cactus: Chemical, nutraceutical composition and potential bio‐ pharmacological properties. Phytother Res. 2020;35(3):1248-1283.
- Available:https://doi.org/10.1002/ptr.6889D 19. Lima-Nascimento AM, Bento-Silva JS, de Lucena CM, de Lucena RFP. Ethnobotany

of native cacti in the northeast region of Brazil: Can traditional use influence availability? Minutes bot. bras. 2019;33 (2):50–359.

Available:https://doi.org/10.1590/0102- 33062019abb0166

- 20. Gheribi R, Khwaldia K. Cactus mucilage for food packaging applications. Coatings. 2019;9:1-19. Available:https://doi.org/10.3390/coatings9 100655
- 21. Andrade CTS, Marques JGW, Zappi DC. Medicinal use of cacti by Bahian country people. Rev Bras Plantas Med. 2006b; 8(3):36-42.
- 22. Shetty AA, Rana M, Preetham S. Cactus: a medicinal food. J. Food Sci. Technol. 2012;49(5):530–536. Available:https://doi.org/10.1007/s13197- 011-0462-5
- 23. Mizrahi Y. Cereus peruvianus (Koubo) new cactus fruit for the world. Rev. Bras. Frutic. two014;36(1):68-78. Available:https://doi.org/10.1590/0100- 2945-447/13
- 24. Silva SÉ, de Oliveira AJB, da Silva MDFP, Mangolin CA, Gonçalves RAC. Cereus hildmannianus (K.) Schum. (Cactaceae): Ethnomedical uses, phytochemistry and biological activities. J. Ethnopharmacol. 2021;264:113339. Available:https://doi.org/10.1016/j.jep.2020 .113339
- 25. Agostini-Costa ST. Bioactive compounds and health benefits of Pereskioideae and Cactoideae: A review. Food Chem. 2020; 327:126961. Available:https://doi.org/10.1016/j.foodche m.2020.126961
- 26. Meiado MV, Machado MC, Zappi DC, Taylor NP, Siqueira FJA. Ecological Attributes, Geographic Distribution and Endemism of Cacti From the São Francisco Watershed. Gaia Scientia. 2015; 9(2):40-53. Available:https://periodicos3.ufpb.br/index.

php/gaia/article/view/24128

27. IUCN - International Union for the Conservation of Nature. Red list of threatened species. Accessed 02 March 2024.

Available: http://www.iucnredlist.org

28. Santos-Díaz MS, Camarena-Rangel NG. Cacti for production of metabolites: Current state and perspectives. Appl. Microbiol. Biotechnol. 2019;103:8657–8667.

Available:https://doi.org/10.1007/s00253- 019-10125-5

- 29. Santana MCD, Santos PAA, Ribeiro ADS. Levantamento etnobotânico da família Cactaceae no estado de Sergipe. Revista Fitos. 2018;12(1):41-53. Available:https://doi.org/10.5935/2446- 4775.20180005
- 30. Betancourt C, Cejudo-Bastante MJ, Heredia FJ, Hurtado N. Pigment composition and antioxidant capacity of betacyanins and betaxanthins fractions of Opuntia dillenii (Ker Gawl) Haw cactos fruit. Food Res. Int. 2017;101:173–179. Available:https://doi.org/10.1016/j.foodres. 2017.09.007
- 31. Lanuzza F, Occhiuto F, Monforte MT, Tripodo MM, Angelo VD, Galati EM. Antioxidant phytochemicals of *Opuntia ficus-indica* (L.) Mill. cladodes with potential anti-spasmodic activity. Pharmacogn. Mag. 2017; 13(Suppl 3): S424–S429. Available:https://doi.org/10.4103/pm.pm_4

95_16

32. Wright CR, Setzer WN. Chemical composition of volatiles from Opuntia littoralis, Opuntia ficus-indica and Opuntia prolifera growing on Catalina Island, California. Nat. Prod. Res. 2014;28(3): 208–211. Available:https://doi.org/10.1080/14786419

.2013.867345

- 33. Benattia FK, Arrar Z, Dergal F. Chemical composition and nutritional analysis of seeds cactus (Opuntia ficus-indica. L). Curr. Nutr. Food Sci. 2019;15:394–400. Available:https://doi.org/10.2174/15734013 14666171228151651
- 34. Štarha R, Chybidziurová A, Lacný Z. Constituents of Turbinicarpus alonsoi Glass & Arias (Cactaceae). Acta Univ. Palacki. Olomuc. 1999;38: 71–73.
- 35. Bruhn JG, Bruhn C. Alkaloids and ethnobotany of mexican peyote cacti and related species. Econ. Bot. 1973;27:241– 251.
- 36. Schwarz A, Medeiros I, Mourão C, Queiroz F, Pugmacher S. Phytochemical and toxic analysis of an ethanol extract from Cereus jamacaru. Toxicol. Lett. 2010;196:S344. Available:https://doi.org/10.1016/j.toxlet.20 10.03.1089.
- 37. Davet A, Carvalho JLS, Dadalt RC, Vituoso S, Dias JFG, Miguel MD, Miguel OD. Cereus jamacaru: a non bu{ered LC quantication method to nitrogen

compounds. Chroma. 2009; 69(S2):245– 247.

Available:https://doi.org/10.1365/s10337- 009-1130-z.

- 38. Medeiros IU, Medeiros RA, Bortolin RH, Queiroz FM, Silbiger VN, Pflugmacher S, Schwarz A. Genotoxicity and pharmacokinetic characterization of Cereus jamacaru ethanolic extract in rats. Biosci. Rep. 2019;39(1):BSR20180672. Available:https://doi.org/10.1042/BSR2018 0672
- 39. Santos JF, Goncalves JLC, Jesus RA, Lima NPC, Scher R, de Oliveira Junior AM, da Silveira Moreira JDJ. Bioactive pro^le of mandacaru fruits and cytotoxicity against the L929 cell line. J. Med. Plant Re. 2021; 15(5):215–225. Available:https://doi.org/10.5897/JMPR202 0.7060.
- 40. Melgar B, Dias MI, Ciric A, Sokovic M, Garcia-Castello EM, Rodriguez-Lopez AD. By-product recovery of Opuntia spp. peels: Betalainic and phenolic profiles and bioactive properties. Ind Crops Prod. 2017; 107:353-359. Available:https://doi.org/10.1016/j.indcrop.

2017.06.011

41. Lee EH, Kim HJ, Song YS, Jin C, Lee KT, Cho J, Lee YS. Constituents of the stems and fruits of Opuntia ficus-indica var. saboten. Arch Pharm Res. 2003;26(12): 1018–1023.

Available:https://doi.org/10.1007/BF02994 752

- 42. Thi Tran TM, Nguyen Thanh B, Moussa-Ayoub TE, Rohn S, Jerz G. Profiling of polar metabolites in fruits of Opuntia stricta var. dillenii by ion-pair high-performance countercurrent chromatography and offline electrospray mass-spectrometry injection. J Chromatogr A; 2019. Available:https://doi.org/10.1016/j.chroma. 2019.06.009
- 43. Silva-Barbosa A, Goodger JQD, Woodrow IE, Pereira de Andrade A, Alcântara-Bruno RL, Souza-Aquino I. Elucidation of the betalainic chromoalkaloid profile of Pilosocereus catingicola (Gürke) Byles & Rowley subsp. Salvadorensis (Werderm.) Zappi (Cactaceae) from Paraíba, Brazil. Afr J Agric Res. 2027;12:1236–1243. Available:https://doi.org/10.5897/AJAR201 6.11019
- 44. Kaur G, Thawkar B, Dubey S, Jadhav P. Pharmacological potentials of betalains. J. Complement. Integr. Med. 2018;15:1–9.

Available:https://doi.org/10.1515/jcim-2017-0063

- 45. Hussain EA, Sadiq Z, Zia-Ul-Haq M. Betalains: Biomolecular aspects. Suiça: Springer International Publishing; 2018.
- 46. García-Cruz L, Dueñas M, Santos-Buelgas C, Valle-Guadarrama S, Salinas-Moreno Y. Betalains and phenolic compounds profiling and antioxidant capacity of pitaya (Stenocereus spp.) fruit from two species (S. Pruinosus and S. stellatus). Food Chem. 2017;234:111–118. Available:https://doi.org/10.1016/j.foodche m.2017.04.174
- 47. Fathordoobady F, Manap MY, Selamat J, Singh AP. Development of supercritical fluid extraction for the recovery of betacyanins from red pitaya fruit (Hylocereus polyrhizus) peel: a source of natural red pigment with potential antioxidant properties. Int. Food Res. J. 2019;26:1023–1034.
- 48. Wybraniec S, Nowak-Wydra B. Mammillarinin: A new malonylated betacyanin from fruits of Mammillaria. J. Agric. Food Chem. 2007;55(20):8138– 8143.

Available:https://doi.org/10.1021/jf071095s 49. Gonçalves ASM, Peixe RG, Sato A,

Muzitano MF, de Souza ROMA, de Barros MT. Pilosocereus arrabidae (Byles & Rowley) of the Grumari sandbank, RJ, Brazil: Physical, chemical characterizations and antioxidant activities correlated to detection of flavonoids. Food Res. Int. 2015;70:110–117. Available:https://doi.org/10.1016/j.foodres.

2014.10.009

- 50. Mena P, Tassotti M, Andreu L, Nuncio-Jáuregui N, Légua P, Del Rio D, Hernández F. Phytochemical characterization of diferente prickly pear (*Opuntia ficus-indica* (L.) Mill.) cultivars and botanical parts: UHPLC-ESI-MSn metabolomics profiles and their chemometric analysis. Food Res. Int. 2018;108:301–308. Available:https://doi.org/10.1016/j.foodres. 2018.03.062
- 51. Osorio-Esquivel O, Ortiz-Moreno A, Garduño-Siciliano L, Álvarez VB, Hernández-Navarro MD. Antihyperlipidemic effect of methanolic extract from Opuntia joconostle seeds in mice fed a hypercholesterolemic diet. Plant Foods Hum. Nutr. 2012;67:365–370.

Available:https://doi.org/10.1007/s11130- 012-0320-2

- 52. Cha MN, Jun HI, Lee WJ, Kim MJ, Kim MK, Kim YS. Chemical composition and antioxidant activity of Korean cactus (Opuntia humifusa) fruitFood Sci. Biotechnol. 2013;22:523-529.
- 53. Astello-García MG, Cervantes I, Nair V, del Socorro SDM, Reyes-Agüer A, Guéraud F. et al. Chemical composition and phenolic compounds profile of cladodes from Opuntia spp. cultivars with different domestication gradient. J. Food Compos. Anal. 2015;43:119–130.
- 54. Ferreres F, Grosso C, Gil-Izquierdo A, Valentão P, Mota AT, Andrade PB. Optimization of the recovery of high-value compounds from pitaya fruit by products using microwave-assisted extraction. Food Chem. 2017;230:463–474. Available:https://doi.org/10.1016/j.foodche m.2017.03.061
- 55. Ammar I, Salem MB, Harrabi B, Mzid M, Bardaa S, Sahnoun Z, et al. Anti-
inflammatory activity and phenolic inflammatory activity composition of prickly pear (Opuntia ficusindica) flowers. Ind. Crops Prod. 2018;112: 313–319. Available:https://doi.org/10.1016/j.indcrop.

2017.12.028

- 56. Kim HK, Tan CP, Karim R, Ariffin AA, Bakar J. Chemical composition and DSC thermal properties of two species of Hylocereus cacti seed oil: Hylocereus undatus and Hylocereus polyrhizus. Food Chem. 2010;119(4):1326–1331. Available:https://doi.org/10.1016/j.foodche m.2009.09.002
- 57. Cabañas-García E, Areche C, Jáuregui-Rincón J, Cruz-Sosa F, Pérez-Molphe BE. Phytochemical profiling of Coryphantha macromeris (Cactaceae) growing in greenhouse conditions using ultra-highperformance liquid chromatography– tandem mass spectrometry. Molecules. 2019;24(4):705. Available:https://doi.org/10.3390/molecules

24040705

58. Salt TA, Tocker JE, Adler JH. Dominance of Δ5-sterols in eight species of the Cactaceae. Phytochem. 1987;26(3):731– 733. Available:https://doi.org/10.1016/S0031- 9422(00)84774-3

59. Jiang J, Li Y, Chen Z, Min Z, Lou F. Two novel C29-5β-sterols from the stems of Opuntia dillenii. Steroids. 2006;71:1073– 1077

- 60. Chahdoura H, Barreira JCM, Barros L, Santos-Buelga C, Ferreira ICFR, Achour L.
Phytochemical characterization and characterization and antioxidante activity of Opuntia microdasys (Lehm.) Pfeiff flowers in diferente stages ofmaturity. J. Food Funct. 2014;9:27–37. Available:https://doi.org/10.1016/j.jff.2014. 04.011
- 61. Núñez-Gastélum JA, González-Fernández R, Hernández HÁ, Campas-Baypoli ON, Rodríguez-Ramírez R, Lobo-Galo N, Valero-Galván J. Morphological characteristics, chemical composition and antioxidant activity of seeds by four wild Opuntia species from north of Mexico. J. Prof. Assoc. Cactus Dev. 2018;20:23–33.
- 62. Kinoshita K, Koyama K, Takahashi K, Kondo N, Yuasa H. New triterpenes from Trichocereus bridgesii. J. Nat. Prod. 1992; 55(7):953–955. Available:https://doi.org/10.1021/np50085a 017
- 63. Ye Y, Kinoshita K, Koyama K, Takahashi K, Kondo N, Yuasa H. New triterpenes from Machaerocereus eruca. J. Nat. Prod. 1988;61(4), 456–460. Available:https://doi.org/10.1021/np970364 d
- 64. Kakuta K, Baba M, Ito S, Kinoshita K, Koyama K, Takahashia K. New triterpenoid saponins from cacti and anti-type I allergy activity of saponins from cactus. Bioorg. Med. Chem. Lett. 2012;22:4793–4800. Available:https://doi.org/10.1016/j.bmcl.201 2.05.058
- 65. Sri Nurestri AM, Sim KS, Norhanom AW. Phytochemical and cytotoxic investigations of Pereskia grandifolia Haw. (Cactaceae) leaves. J. Biol. Sci. 2009;9:488–493. Available:https://doi.org/10.3923/jbs.2009. 488.493
- 66. Ciriminna R, Delisi R, Albanese L, Meneguzzo F, Pagliaro M. Opuntia ficusindica seed oil: Biorefinery and bioeconomy aspects. Eur. J. Lipid Sci. Technol. 2017;119: 1700013. Available:https://doi.org/10.1002/ejlt.20170 0013
- 67. Imai T, Okazaki S, Kinoshita K, Koyama K, Takahashi K, Yuasa H. Triterpenoid saponins from cultural plants of Stenocereus stellatus (Cactaceae). J. Nat. Med. 2006;60:49–53.
- 68. Okazaki S, Kinoshita K, Koyama K, Takahashi K, Yuasa H. New triterpene saponins from Stenocereus eruca (Cactaceae). J. Nat. Med. 2007;61:24–29.
- 69. Murillo E, Melendez-Martinez A, Portugal F. Screening of vegetables and fruits from Panama for rich sources of lutein and zeaxanthin. Food Chem. 2010;122(1):167– 172.

Available:https://doi.org/10.1016/j.foodche m.2010.02.034

70. Martin AA, Freitas RA, Sassaki GL, Evangelista PHL, Sierakowski MR. Chemical structure and physical-chemical properties of mucilage from the leaves of Pereskia aculeata. Food Hydrocolloids. 2017;70:20–28. Available:https://doi.org/10.1016/j.foodhyd. 2017.03.020

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