

Journal of Complementary and Alternative Medical Research

Volume 25, Issue 4, Page 31-39, 2024; Article no.JOCAMR.115406 ISSN: 2456-6276

Modulatory Functions of Craterispermum schweinfurthi on the Hypothalamic-Pituitary-Gonadal Axis of Male Wistar Rats in Phenyl Hydrazine Induced Testicular Toxicity

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Authors' contributions

This work was carried out in collaboration among all authors. Authors SF and PA conceptualized and designed the study. Author SF conducted the experiments and collected the data. Authors DJD and AJE performed the statistical analysis and wrote the manuscript. Authors SF, DJD, AJE and PA critically reviewed and edited the manuscript. All authors contributed to the interpretation of results and approved the final version for submission. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JOCAMR/2024/v25i4530

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/115406

> Received: 05/02/2024 Accepted: 10/04/2024 Published: 16/04/2024

Original Research Article

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J. Compl. Altern. Med. Res., vol. 25, no. 4, pp. 31-39, 2024

ABSTRACT

Introduction: Modulation of the hypothalamic-pituitary-gonadal axis **is** mediated by different factors which are of research interest.

Aim: To evaluate the modulatory functions of *Craterispermum schweinfurthi* leaf extract on the hypothalamic-pituitary-gonadal axis of male Wistar rats in phenyl hydrazine induced testicular toxicity.

Methodology: 40 male Wistar rats weighing between 100-250g were randomly divided into 8 groups of 5 rats each. Testicular toxicity was induced through intraperitoneal administration of 40mg/kg of phenyl hydrazine at 9am on day 0 and two additional injections at 9am and 6pm on day 1 in all rat groups except groups 1 and 8 and were treated as follows for 14 days; Group 1: Rats in this group received distilled water only: Group 2: Untreated Phenyl hydrazine induced toxicity rats: Groups 3-5 received 250mg/kg, 500mg/kg and 750mg/kg body weight of the extract: Group 6: Rats in this group were administered 0.23ml/kg of Bioferon: Group7: Phenyl hydrazine + Phytosterol (2000mg/kg): Group 8: Phytosterol only (2000mg/kg). 24 hours after the last administration, the rats were anaesthetized using 3.5% chloroform soaked in cotton wool and blood samples collected through direct cardiac puncture for the estimation of serum concentration of reproductive hormones. Also, rat's caudal epididymis was excised for the determination of sperm indices.

Results: Administration of the hydromethanol leaf extract of *Craterispermum schweinfurthi* to rats Groups 3-5, significantly increased serum concentration of luteinizing, Follicle stimulating hormones and Testosterone compared to Group 2 (phenyl hydrazine induced toxicity) rats (p<0.05): Suggesting a possible modulatory function of the extract. Significantly dose dependent higher values of sperm volume, viability, count, normal and active sperm were observed amongst groups 3-5 rats following the administration of graded doses of the extract compared to Group 2 (phenyl hydrazine induced toxicity) rats (p<0.05). Suggesting a possible amelioration of the toxic effects of phenyl hydrazine.

Conclusion: This study reports that administration of hydromethanol extract of C*raterispermum schweinfurthi* caused a significant and dose dependent improvement in the concentration of male reproductive hormones: resulting in a predictable increase in sperm indices.

Keywords: Craterispermum schweinfurthi; Phenyl hydrazine; toxicity; hypothalamic-pituitary-gonadal axis.

1. INTRODUCTION

A challenging global phenomenon affecting in adequate understanding, mankind lies prevention, management and treatment of the ever-increasing male infertility, infertility is defined as the inability to conceive after about a year of unprotected regular sexual intercourse [1]. With about 12% prevalence rate, male infertility impacts over 30 million people globally. Male infertility is a major contributing factor to about 30% or more reported cases of infertility worldwide [2-3]. Male fertility depends largely on the serum concentration of male conceptive hormones: Luteinizing hormone, Follicle stimulating hormone & Testosterone and sperm characteristic: sperm count, quality, motility, viability, morphology, defects in any of these factors can cause infertility [4] About 90% of all reported infertile cases have a direct association with hormonal and sperm indices [5]. Elevated temperature. endocrine disorders. scrotal lifestyle, environmental and nutritional factors

have all been reported to negatively impact sperm parameters resulting in male infertility [6]. Most of the aforementioned factors responsible for male infertility can be reversed surgically or therapeutically using drugs [7]. However, treatment options solely depend on the possible cause of male infertility, financial status, facilities available in a designated hospital, the patient's age and expertise [8].

In recent years, complementary therapies for infertility have received growing attention, and various nutritional approaches, and medicinal plants have been explored for the treatment of male reproductive disorders [9]. Several local medicinal plants with fertility boosting effects have been traditionally used globally [10-11]. Fertility-related properties of plants are also of interest in modern day scientific research [12]. Specific important compounds identified in most medicinal plants are effective in the treatment, management, and prevention disease of conditions [13]. Contemporary approaches to infertility treatment globally have received growing attention following men's increasing and reliance on effective herbal interest supplementation [14]. The European Association of Urology and the World Health Organization (WHO) have recently reported the use of traditional medicine as a multidimensional integrative approach to infertility treatment [15-16]. Such a growing interest in medicinal plants including Craterispermum schweinfurthi has inspired scientists to clarify their effects in fertility studies as such interventions would serve as possible beneficial alternatives to mankind against the alreadv existina orthodox Craterispermum schweinfurthi medications. species are shrubs with axillary paired at the nodes and often condensed. Its applications in traditional medicine are numerous. In traditional folklore medicine the seed, leaves, and inner bark have been described to have beneficial effects in stomach afflictions, ulcer, infertility, anemia, diabetes and fever [17]. Despite the wide use of Craterispermum schweinfurthi in folklore medicine in our environment, scientific studies on its fertility properties are relatively scanty.

Hence, on account of its many described anecdotal benefits, the present study attempts an evaluation of the potential modulatory functions of the hvdromethanol leaf extract of Craterispermum schweinfurthi on the hypothalamic-pituitary-gonadal axis of male wistar rats in phenyl hydrazine induced testicular toxicity. This is with a view to validating the anecdotal use of the leaf of Craterispermum schweinfurthi as an enhancer of reproductive health in our environment. Also, an attempt was made to compare the effects of phytosterol: a major inherent bioactive compound identified in schweinfurthi Craterispermum leaves [13] against that of Craterispermum schweinfurthi extract.

2. MATERIALS AND METHODS

2.1 Collection, Identification and Extraction of Plant Materials

Fresh leaves of Craterispermum schweinfurthii were obtained from the University of Port Harcourt Botanical Garden. Dr. Chimezie Ekeke of the Department of Plant Science and University Port-Harcourt, Biotechnology, of identified Nigeria and authenticated the specimen and assigned a reference code; UPH/V/296. Voucher specimen was subsequently deposited in the University

Herbarium for future reference. The plant leaves were gathered, and all extraneous materials carefully removed. The leaves were air dried at room temperature for a minimum of 7 days after which it was pulverized into powder and the weighed quantity of 670.6g dissolved using Soxhlet device in 390ml of water-methanol mixture (25:75% v/v BDH) for three days in a jar. It was filtered and concentrated using a rotary evaporator at 40°C and the yield was 73%. Obtained extract was preserved in airtight containers and stocked at room temperature prior administration.

2.2 Procurement and Handling of Experimental Animals

Wistar rats weighing between 100-250g were used for the study. Animals were acquired from the Department of Physiology Animal House, Faculty of Basic Medical Sciences, College of Health Sciences, University of Port Harcourt, Nigeria. Rats were placed in different compartments, one for each experimental group cared for under standard laboratory and conditions. Wood shavings and beddings were changed on a daily basis to prevent any infection due to unkept beddings. The animals were acclimatized for two weeks and subsequently grouped for the study.

2.3 Acute Toxicity Studies

The acute toxicity of the hydromethanol extract of *Craterispermum schweinfurthi* leaves was determined using Karber's method as modified by Aliu and Nwude, (1982) [18]. Lethal dose (LD50) of the extract was found to be 3968mg/kg body weight. The study was conducted in accordance with the guidelines for the care and use of laboratory animals] [19].

2.4 Phenyl hydrazine (PHZ), Drug and Phytosterol Purchase

Phenyl hydrazine (PHZ) was purchased from JHD Co., LTD, 618, Qingshan Road, Licang Dist., Qingdao, Shandong, China; Bioferon procured from Biopharm Quality and Tradition, 12 Klemenova Dacha Street, Apt. 11, Kharkiv, 61033, Ukraine while Phytosterol was obtained from Wakunaga of America Co., LTD. Mission Viejo, CA92691 U.S.A.

2.5 Experimental Design

40 Wistar rats weighing between 100-250g were used for the study. After 14 days of

acclimatization, the rats were randomly divided into 8 groups of 5 rats each: designated Groups 1-8. Phenyl hydrazine testicular toxicity was induced intraperitoneally following 40mg/kg body weight of phenyl hydrazine administration on day 0 and two additional injections at 9am and 6pm on day 1 in all rat groups except groups 1 and 8 as was described previously [20-21]. And were treated as follows for 14 days;

Group 1: Control group; rats in this group received extract vehicle only

Group 2: Untreated Phenyl hydrazine toxicity rats

Group 3: Low extract dose group; rats in this group received 250mg/kg of the leaf extract of Craterispermum schweinfurthi

Group 4: Medium extract dose group; rats in this group received 500mg/kg of the leaf extract of Craterispermum schweinfurthi

Group 5: High extract dose group; rats in this group were given 750mg/kg of the leaf extract of Craterispermum schweinfurthi

Group 6: Bioferon group; rats in this group were administered 0.23ml/kg of Bioferon [22,21].

Group7: Phenyl hydrazine toxicity + Phytosterol (2000mg/kg)

Group 8: Phytosterol only (2000mg/kg)

24 hours after the last administration, the rats were anaesthetized using 3.5% chloroform soaked in cotton wool and blood samples collected through direct cardiac puncture and immediately transferred into plain sample tubes for the estimation of serum concentration of reproductive hormones: Luteinizing hormone, Follicle stimulating hormone and Testosterone. Also, rats caudal epididymides containing sperm were excised for the determination of sperm indices: Count, Viable, Active, Normal, Abnormal, Volume, Appearance/Morphology etc. The samples were immediately used for the estimation of the above variables.

2.6 Determination of Reproductive Hormones and Sperm Indices

Serum level of luteinizing hormone (LH), follicle stimulating hormone (FSH), and testosterone

was estimated using enzyme immunoabsorbent assay kits (Accu-Bind ELISA Microwells, California, USA). Procedure was as specified in the available manual.

using indices was analyzed Semen the computer-assisted semen analysis (CASA) version 11 (Hamilton Thorne Bioscience). The testes were excised and caudal epididymis carefully isolated and placed in a petri dish containing 3 ml of sodium bicarbonate (NaHCO3) buffered tyrodes' solution. 1 mm incisions were made on them and sperm carefully drawn into a plastic pipette which was subsequently transferred into 5 ml test tubes and shaken for homogeneity/dispersal of sperm cells. The following Sperm indices were evaluated: sperm motility, morphology, count, viability, activeness, sluggishness, dead rate etc.

2.7 Statistical Analysis

Results are as presented in Tables 1 and 2 as Mean ± Standard Error of Mean (SEM). Significant differences were determined using one-way ANOVA and LSD Post Hoc test. A p value of less than 0.05 was considered statistically significant.

3. RESULTS

3.1 Values of Male Reproductive Hormones in Phenyl Hydrazine Induced Toxicity Treated with Extract and Phytosterol

Table 1 shows that significantly lower values of Follicle Luteinizina hormone. stimulating hormone and Testosterone were observed amonast Group 2 rats following the administration of 40mg/kg body weight of Phenyl hydrazine compared to Group 1 (Control) rats Suggesting a possible harmful (p<0.05). reproductive effect of phenyl hydrazine in male Wistar rat. However, upon the administration of doses (250mg/kg, 500ma/ka araded and 750mg/kg) of the extract of Craterispermum schweinfurthi to rats in Groups 3, 4 and 5, significantly higher values of luteinizing hormone, Follicle stimulating hormone and Testosterone were observed compared to Group 2 (Untreated phenyl hydrazine) rats (p<0.05). Indicating a possible modulatory function of the extract in male Wistar rats. Surprisingly, the values of these hormones were significantly increased (p<0.05) in a dose dependent manner with the

administration of the extract. Similarly, Bioferon administration to Group 6 rats shows a significant improvement in the serum concentration of luteinizing hormone, follicle stimulating hormone and testosterone compared to Group 2 rats (p<0.05). At a dose of 750mg/kg, the extract exhibited an increase of 0.36±0.004, 0.20±0.003 and 0.62±0.003 respectively in luteinizing hormone, Follicle stimulating hormone and Testosterone compared to Bioferon with an increase of 0.32±0.003, 0.18±0.005 and 0.61±0.003 in luteinizing hormone, Follicle stimulating hormone and Testosterone. Suggesting a possible greater potency of the extract at 750mg/kg body weight.

Also, Groups 7 and 8 rats administered 2000mg/kg body weight of phytosterol shows significantly higher values of luteinizing hormone, Follicle stimulating hormone and Testosterone compared to Group 2 rats (p<0.05).

3.2 Values of Sperm Indices in Phenyl Hydrazine Induced Toxicity Treated with Extract and Phytosterol

Table 2 shows significant reduction in sperm volume, viability, count, active and normal sperms amongst Group 2 rats following phenyl hydrazine administration compared to Group 1

(Control) rats. Also, there was a corresponding and significant increase in the population of sperms that were, abnormal, sluggish and dead compared to Group 1 rats. Indicating a possible harmful effect of phenyl hydrazine on sperm indices in male Wistar rats. Administration of graded doses of the extract of Craterispermum schweinfurthii to Groups 3-5 rats demonstrated a dose dependent significant improvement in sperm volume, viability, count, normal and active sperms compared to Group 2 (phenyl hydrazine induced toxicity) rats (p<0.05). However, sperm viscosity and appearance remained unchanged thought out the duration of the study. Population of abnormal, sluggish and dead sperms were significantly decreased amongst Groups 3-5 rats compared to Group 2 (p<0.05). These findings are indicative of a possible beneficial effects of the extract on sperm parameters. Significant increases in sperm viability, count, active and normal sperms were also observed following Bioferon administration to rats in group 6 compared to Group 2 rats (p<0.05). Similarly, phytosterol administration to Groups 7 and 8 rats caused a significant increase in sperm viability, count, normal and active sperm while the population of abnormal, sluggish and dead sperms were significantly decreased compared to Group 2 rats: suggesting a possible reversal of the deleterious effect of phenyl hydrazine in male Wistar rats.

Table 1. Values of male reproductive hormones in phenyl hydrazine induced toxicity treatedwith extract and phytosterol

	Groups	Luteinizing Hormone (miu/ml)	FollicleStimulating Hormone (miu/ml)	Testosterone (ng/ml)
1	Control	0.30±0.005 ^b	0.15±0.003 ^b	0.58±0.003 ^b
2	Untreated Phenyl hydrazine toxicity rats	0.25±0.004ª	0.10±0.000 ª	0.45±0.004 ^a
3	Phenyl hydrazine + 250mg/kg Extract	0.29±0.002 ^b	0.14±0.003 ^{ab}	0.54±0.003 ^{ab}
4	Phenyl hydrazine + 500mg/kg Extract	0.34±0.003 ^{ab}	0.19±0.003 ^{ab}	0.60±0.003 ^{ab}
5	Phenyl hydrazine + 750mg/kg Extract	0.36±0.004 ^{ab}	0.20±0.003 ^{ab}	0.62±0.003 ^{ab}
6	Phenyl hydrazine + Bioferon	0.32±0.003 ^{ab}	0.18±0.005 ^{ab}	0.61±0.003 ^{ab}
7	Phenyl hydrazine + 2000mg/kg Phytosterol	0.28±0.003 ^{ab}	0.14±0.003 ^{ab}	0.50±0.003 ^{ab}
8	2000mg/kg Phytosterol only	0.33±0.003 ^{ab}	0.17±0.003 ^{ab}	0.60±0.003 ^{ab}

Values are shown as Mean ± SEM; n=5; ^a Significant at P<0.05 compared with Group 1 (control). ^b Significant at p<0.05 compared with Group 2 (untreated phenyl hydrazine induced toxicity).

Table 2. Values of sperm indices	in phenyl ł	vdrazine induced toxicit	y treated with extract and phytosterol
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	Control	Untreated Phenyl hydrazine toxicity rats	Phenyl hydrazine + 250mgkg Extract	Phenyl hydrazine + 500mg/kg Extract	Phenyl hydrazine + 750mg/kg Extract	Phenyl hydrazine + Bioferon	Phenyl hydrazine + 2000mg/kg Phytosterol	2000mg/kg Phytosterol only
Volume (ul)	0.2.00±0.001 ^b	0.1.00±0.002 ^a	0.2.00±0.001 ^b	0.2.00±0.000 ^b	0.3.00±0.008 ^{ab}	0.2.00±0.000 ^b	0.2.00±0.001 ^b	0.2.00±0.000 ^b
Ρh	8.00±0.002	8.00±0.001	8.00±0.000	8.00±0.000	8.00±0.001	8.00±0.000	8.00±0.002	8.00±0.002
Viability (%)	90.00±0.002 ^b	70.00±0.007 ^a	70.00±0.001ª	80.00±0.008 ^{ab}	85.00±0.00 ^{ab}	80.00±0.002 ^{ab}	70.00±0.00 ^a	80.00±0.001 ^{ab}
Count	600.00±0.001	400.00±0.00 ^a	500.00±0.002 ab	600.00±0.005 ^b	700.00±0.003 ^{ab}	600.00±0.001 ^b	500.00±0.001 ^{ab}	500.00±0.005 ^{ab}
Normal (%)	80.00±0.000	60.00±0.001ª	70.00±0.002 ^{ab}	75.00±0.001 ^{ab}	80.00±0.00 ^b	75.00±0.000 ^{ab}	70.00±0.005 ^{ab}	80.00±0.002 ^b
Abnormal (%)	20.00±0.001b	35.00±0.002ª	30.00±0.001 ^{ab}	25.00±0.004 ^{ab}	20.00±0.002 ^b	25.00±0.000 ^{ab}	30.00±0.000 ^{ab}	20.00±0.000 ^b
Active (%)	80.00±0.002 ^b	60.00±0.001ª	70.00±0.003 ^{ab}	80.00±0.000 ^b	85.00±0.001 ^{ab}	80.00±0.001 ^b	70.00±0.007 ^{ab}	80.00±0.006 ^b
Sluggish (%)	5.00±0.001 ^b	10.00±0.001ª	10.00±0.000 ^a	10.00±0.002ª	5.00±0.000 ^b	10.00±0.001ª	10.00±0.002ª	10.00±0.001ª
Dead	10.00±0.005 ^b	25.00±0.003 ^a	20.00±0.000 ^{ab}	10.00±0.002 ^b	10.00±0.001 ^b	10.00±0.002 ^b	20.00±0.003 ^{ab}	10.00±0.002 ^b
Appearance	Milky	Milky	Milky	Milky	Milky	Milky	Milky	Milky
Viscosity	Normal	Normal	Normal	Normal	Normal	Normal	Normal	Normal

Values are shown as Mean ± SEM; n=5; ^a Significant at P<0.05 compared with Group 1 (control). ^b Significant at p<0.05 compared with Group 2 (untreated phenyl hydrazine induced toxicity)

4. DISCUSSION

Recent researches on medicinal plants have assumed an incredible global recognition in past vears. The use of some identified plant constituents in pharmaceutical supplementation and intervention have come a long way in the elevation of the status of traditional medicine in West Africa [23]. The need for fertility modulation enhancement in men cannot and be overemphasized. In the present study, obtained results showed a significant improvement and elevation in the serum concentration of luteinizing hormone, follicle stimulating hormone and testosterone in male Wistar rats following the administration of leaf extract of Craterispermum schweinfurthi compared with the control. This suaaest probably that Craterispermum schweinfurthi extract plays an important role in the modulation and improvement of hormonal confers pro-fertility functions. level which Luteinizing hormone stimulates the production of testosterone by the Leydig cells, which causes Sertoli and peritubular cells of the the seminiferous tubules to initiate spermatogenesis [24-25]. Increased secretion of Follicle stimulating hormone aids spermatogenesis, fertility and gonadal development. LH and FSH secreted by the pituitary gland are of major importance in male reproduction. Increased testosterone concentration indicates that the extract improves libido: Testosterone concentration is associated with the gonadotropins such that an increased secretion would predictably induce relative increase in testosterone secretion [26]. Our findings are consistent with Allouh et al. (2015) [27], who reported elevation in serum earlier an reproductive hormones concentration following the administration of medicinal plants with aphrodisiac properties in male Wistar rats.

The modulatory functions of the leaf extract of Craterispermum schweinfurthi on sperm indices was examined in the present study. Nowadays, medicinal plant extracts have been given due recognition and their effects on various organs and tissues of the body identified and documented. Reproductive tissues like testis and epidydimal tissues are major target tissues of plant extracts with aphrodisiac properties. Sperm motility, viability, count etc. are important factors in natural or experimental reproductive functions. In fertile men, sperm indices especially motility, viability and count are directly associated with copulatory potentials [28]. Scientists believe that free radicals in the testicular region are largely

responsible for dvsfunction in sperm characteristics and sperm cell membrane fluidity. cvtoplasmic bridaes which destrovs and ultimately decrease sperm count and motility [29-Apparently, the antioxidant properties 301. of Craterispermum schweinfurthi improved the quality of sperm by increasing the expression of sperm indices and cell membrane stabilization [31]. Findings from this study are in line with Wong et al., 2006 [32] and Oyeyemi, 2008 [4] in which extracts of plants improved sperm indices and oxidative stress.

5. CONCLUSION

This study reports that administration of hydromethanol extract of Craterispermum schweinfurthi caused a significant and dose dependent increase in the concentration of male reproductive hormones: resulting in a predictable improvement in sperm indices in male Wistar rats. The actual mechanism of action is presently unclear and would require further studies.

CONSENT

It is not applicable.

ETHICAL APPROVAL

Ethical approval was sought and obtained from the University of Port Harcourt Ethical Committee vide a communication referenced: UPH/CEREMAD/REC/MM82/024 and dated 23rd November 2021.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Alahmar AT. Role of oxidative stress in male infertility: An Updated Review. J. Hum. Reprod. Sci. 2019;12, 4–18.
- Agarwal A, Roychoudhury S, Bjugstad KB, Cho C-L. Oxidation-reduction potential of semen: what is its role in the treatment of male infertility? Ther Adv Urol. 2016; 8:302–18.
- Hosseini H, Abdi F. Experiences of vasectomy: A phenomenological study. N Am J Med Sci. 2012;4:619–23.
- 4. Oyeyemi MO, Olukole SG, Esan O. Sperm morphological studies of West African

Dwarf Bucks treated with pumpkin plant (Cucurbita pepo). Int J Morphol. 2008; 26:121–6.

- 5. Roozbeh N, Rostami S, Abdi F. A Review on herbal medicine with Fertility and Infertility characteristics in Males. IJOGI. 2016;19:18–32.
- Marbeen MI, AL-Snafi AE, Marbut MM, Allahwerdy IY. The probable therapeutic effects of date palm pollen in the treatment of male infertility. Tikrit Journal Pharm Sci. 2005;1:30–5.
- 7. Abdillahi H, Van Staden J. South African plants and male reproductive healthcare: Conception and contraception. J. Ethnopharmacol. 2012;143:475–480.
- Nwajiaku LA, Mbachu, II, Ikeako, L. Prevalence, Clinical Pattern and Major Causes of Male Infertility in Nnewi, South East Nigeria: A Five-year Review. Afrimedic J. 2012;3:16–19.
- 9. Abdi F, Roozbeh N, Mortazavian AM. Effects of date palm pollen on fertility: research proposal for a systematic review. BMC Res Notes. 2017;10:363.
- Saronee F, Amadi J, Azosibe P. Effects of the Hydromethanol Leaf Extract of *Craterispermum schweinfurthi* on Sperm Characteristics and Hematological Indices in Male Wistar Rats. Cross Curr Inter J Med Biosci. 2024;6(1):21-25.
- Saronee F, Bekinbo MT, Ojeka SO, Dapper DV. Comparative assessment of methanolic extracts of hog plum (*spondias mombin* linn.) leaves and turmeric (*Curcuma longa* L.) rhizomes on blood glucose and glycosylated haemoglobin in male wistar rats. J Appl Sci Environ Mgt, 2019;23(9):1631-1636.
- Jain S, Choudhary GP, Jain DK. Medicinal plants with potential anti-fertility activity: A review. Inter J Green Pharm. 2015;9:223.
- 13. Saronee F, Amah-Tariah FS, Chinko BC, Dapper DV. GC-MS and Proximate Analysis of the Hydromethanol Extract of *Craterispermum schweinfurthi* Leaves. South Asian Res J Nat Prod, 2023;6(2): 101-109.
- 14. Yao DF, Mills JN. Male infertility: Lifestyle factors and holistic, complementary, and alternative therapies. Asian J androl. 2016; 18:410–8.
- 15. Nejatbakhsh F, Shirbeigi L, Rahimi R, Abolhassani H. Review of local herbal compounds found in the Iranian traditional medicine known to optimise male

fertility. Andrologia. 2016;48:850-9.

- 16. Nantia E, Moundipa PF, Monsees TK, Carreau S. Medicinal plants as potential male anti-infertility agents: A review. Androl .2009;19:148–58.
- Sofowora, A. Screening plants for bioactive agents. In medicinal plants and traditional medicine in Africa. Spectrum books limited, Ibadan, 2ed. 1993;134-156.
- Aliu, YO, Nwude N. Veterinary pharmacology and toxicology experiments. Zaria: ABU Press, Zaria; 1982.
- (US) NRC. Committee for the Update of the Guide for the Care and Use of Laboratory Animals; 2019. Available:https://www.ncbi.nlm.nih.gov/boo ks/NBK54050/ downloaded on the 20th of June, 2022.
- 20. Tariq HA. Hematinic and anti-anemic effect of T hymoquinone against phenylhydrazine-induced haemolytic anemia in rats. Res. J. Med. sci. 2014;8 (2):67-72.
- 21. Saronee F, Amah-Tariah FS, Chinko BC, Dapper DV. Ameliorative effects of the hydromethanol leaf extract of *Craterispermum schweinfurthii* on phenyl hydrazine induced anemia in male wistar rats. Inter J Recent Res Life Sci. 2023; 10(3):19-25.
- 22. Zara M, Samuel A, Amina Z. Characterisation of bioactive componenets of *Gossypium barbadense* L. with Hematinic potential in wistar rats. BJPMR. 2014;4(21):2563-2574.
- 23. Edeoga, HO., Okwu, DE, Mbaoble, BO. Phytochemical constituents of some Nigerian Medicinal plants. African J Biotecnol. 2005;4(7):685-688.
- Airaodion, AI, Ogbuagu EO, Ekenjoku JA, Okoroukwu VN, Ogbuagu U, Airaodion EO. Antifertility effect of ethanolic leaf extract of Carica papaya in male Wistar rats. Merit Res J Med Med Sci. 2019; 7(10):374-381.
- 25. Lakhsman J, Changamma C. Antispermatogenic effect of Vernonia amygdalina seed extract on steroidogenesis in albino rats. Inter J Pharm Pharmaceut Sci. 2013;5(1):25-36.
- 26. Jameson JL, De Groot LJ. Endocrinology: Adult and Pediatric E-Book. Elsevier Health Sci. 2015;2: 21-79.
- 27. Allouh MZ, Daradka HM, Ghaida JHA. Influence of Cyperus esculentus tubers (Tiger Nut) on male rat copulatory

behavior. BMC Complement Alter Med. 2015;15(331):1-7.

- Zhou Q, Li Y, Nie R, Friel P, Mitchell D, Evanoff RM. Expression of stimulated by retinoic acid gene 8 (Stra8) and maturation of murine gonocytes and spermatogonia induced by retinoic acid *in vitro*. Biol Reprod. 2008;78:537– 45.
- 29. Aziz N, Saleh RA, Sharma RK, Lewis-Jones I, Esfandiari N, Thomas AJ, Jr. Novel association between sperm reactive oxygen species production, sperm morphological defects, and the sperm deformity index. Fertil Steril. 2004;81:349– 54.
- Kolarovic J, Popovic M, Zlinská J, Trivic S, Vojnovic M. Antioxidant activities of celery and parsley juices in rats treated with doxorubicin. Molecules. 2010;15:6193–20 4.
- Shi TY, Chen G, Huang X, Yuan Y, Wu X, Wu B. Effects of reactive oxygen species from activated leucocytes on human sperm motility, viability and morphology. Andrologia. 2012;44(Suppl 1):696–703.
- Wong PY, Kitts DD. Studies on the dual antioxidant and antibacterial properties of parsley (*Petroselinum crispum*) and cilantro (*Coriandrum sativum*) extracts. Food Chem. 2006;97:505–15.

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