

Synergistic Effect of Phosphine and Carbon Dioxide on the Mortality of *Tribolium castaneum* (Coleoptera: Tenebrionidae) in Paddy

R. Meenatchi¹, R. P. S. J. Alice¹ & P. P. Paulin¹

¹ Indian Institute of Food Processing Technology, Tamil Nadu, India

Correspondence: R. Meenatchi, Indian Institute of Food Processing Technology, Thanjavur, Tamil Nadu-613005, India. Tel: 91-975-096-8420. E-mail: meena@iifpt.edu.in

Received: January 3, 2018

Accepted: March 17, 2018

Online Published: June 15, 2018

doi:10.5539/jas.v10n7p503

URL: <https://doi.org/10.5539/jas.v10n7p503>

Abstract

Phosphine (PH₃) is widely used as a fumigant for the control of stored product pests. Indiscriminate use of PH₃ leads to the development of resistant strains. Thus, an experiment was conducted in a laboratory scale fumigation chamber to determine the synergistic effect of carbon dioxide (CO₂) and PH₃ on the mortality of *Tribolium castaneum* during fumigation of paddy. PH₃ gas generation varies depending upon grain moisture content and dosage of PH₃ used for fumigation treatment. Thus, the maximum PH₃ concentration with respect to saturation time was achieved when paddy were treated with 98% CO₂ + 3 ppm PH₃ at 17% moisture content, leading to a quick mortality of different life stages of *T. castaneum* with minimum LT₅₀ and LT₉₉ value. Among the different stages of *T. castaneum*, pupal stage was more resistant to all the treatments compared to larva and adult stages. PH₃ residues in all the treatments were below the recommended level and it can be considered as safe for consumption.

Keywords: paddy, moisture, phosphine, generation rate, mortality, *T. castaneum*

1. Introduction

India handles more than 200 million tons (MT) of food grains annually. These grains need to be preserved using the available fumigants. Paddy and wheat are commonly stored in warehouses in large quantities and are often infested with *T. castaneum* due to inappropriate storage and handling. To prevent infestation grains are generally fumigated with aluminum phosphide tablets for more than two decades. However, due to unscrupulous and continuous usage of aluminum phosphide resulted in high level pest resistance to phosphine in stored products (Mau et al., 2012). Mixture of CO₂ and PH₃ is considered as a potential fumigant for the management of stored product pests (Leelaja et al., 2007; Valmas & Ebert, 2006). Carbon dioxide (CO₂) a potential gaseous fumigant toxic to insects at high concentration and requires large exposure time for achieving mortality of all the stages of insects (Hasan et al., 2016). Many studies show that the addition of CO₂ to PH₃ enhances the toxicity of PH₃ and reduces the dose required to kill insects (Ren et al., 1994). The combination of CO₂ with PH₃ increases the penetration rate of the fumigant, acts as the propellant for delivering PH₃, enhances the respiration rate of insects and is more effective than existing solid formulations available in the market (Leesch, 1992). A combination of CO₂ and PH₃ is available in the trade name of ECO₂FUME® and are used in countries such as Australia, New Zealand, North America and Europe (Tumaming et al., 2012). It consists of cylinderized compressed gas formulation containing a mixture of 2% PH₃ by weight (2.6% by volume) in CO₂ (98% by weight). Another recently introduced fumigant for the management of stored product pest is VAPORPH₃OS (99.3 % pure PH₃ mixed with air). These formulations are considered to be safe, effective and are user friendly for fumigating food and non-food commodities (Meenatchi & Alagusundaram, 2014). But due to stringent rules in importing and the non-availability of any other chemicals for fumigation other than PH₃, none of the fumigants were registered in India. At this juncture, a study was undertaken by using PH₃ and CO₂ combinations to evaluate the synergistic effect of PH₃ with respect to saturation time, PH₃ generation in paddy and mortality of *T. castaneum*.

2. Materials and Methods

2.1 Conditioning Paddy

The paddy was procured with the initial moisture content of 12.5% wet basis (w.b.) and conditioned to the required moisture content of 11, 14 and 17%. To obtain lower moisture content, samples were spread in thin

layer under shade for natural air drying for about 8 h. To increase the moisture content, samples were conditioned by adding required amount of distilled water and mixed well by passing through a screw conveyor. The amount of water to be added to increase grain moisture content was calculated by using the following Equation (1). The conditioned grain samples were stored in sealed polythene bags in a deep freezer at -5 ± 2 °C.

Amount of water added,

$$Kg = W_s X \frac{M_f - M_i}{100 - M_f} \quad (1)$$

Where, W_s : Sample weight (Kg); M_f : Final moisture content (% w.b.); M_i : Initial moisture content (% w.b.)

2.2 Culturing of Test Insects

T. castaneum was cultured and maintained at the Storage Engineering Laboratory at IIFPT, Thanjavur. Insects were cultured at 30 °C and 70% RH. Different stages of insects such as pupa, larva and adults were maintained separately to carry out mortality studies.

2.3 Lab Model Fumigation Set up for Toxicity Studies

Phosphine fumigation lab model set up was designed and fabricated at Indian Institute of Food Processing Technology (IIFPT). It consists of a circular outer acrylic cylindrical tube with the dimension of 37×27 cm (ht \times \emptyset) which is pasted in a flat acrylic at the bottom (Figure1). Inside the fumigation chamber, four compartments were made by partitioning it with wire mesh and were fitted in a stand. Fumigation cups of 4×6 cm (ht \times \emptyset) were made by cutting small acrylic tubes and were pasted with fine wire mesh at the bottom. The top of the fumigation setup was covered with PVC end cap to make it air tight. Rubber septum was placed at the top cover for injecting PH_3 . Provisions were made for injecting and releasing CO_2 from the cylinder (Meenatchi et al., 2015). Required volume of PH_3 gas was calculated and injected into the fumigation chamber using Hamilton syringe through the rubber septum located at the top of the fumigation chamber. The 98% CO_2 in air was continuously flowed through the chamber.



Figure 1. Acrylic fumigation chamber

2.4 Measurement of PH_3 Concentration

Phosphine concentration was measured using 250 PM+ portable phosphine monitor (Uniphos Envirotronic Pvt Ltd, India) at every 4 h interval for a period of 120 h.

2.5 Bioassay Studies

Fifteen grams of the conditioned paddy (11, 14 and 17% moisture content) were taken in fumigation cups and 10 insects of various life stages were released. The fumigation cup was covered with muslin cloth. After treatment, the insects were transferred into paddy and then kept in environmental chamber maintained at 25 °C and 60% RH. The number of different life stages of dead *T. castaneum* was recorded after fumigation at different time intervals and the mortality percentage was calculated.

In addition to PH₃ generation and saturation time, a similar set of experiments were carried out to know the effect of various combinations of fumigant on insect mortality at different time intervals. In this study, 98% CO₂ with 3, 2, and 1 ppm PH₃ and 3 ppm PH₃ alone was used as a control. From this data LT₅₀ and LT₉₉ value of various life stages of *T. castaneum* were calculated. All experiments were repeated thrice.

2.6 Determination of PH₃ Residues in the Fumigated Paddy

The fumigated paddy was placed in a tray for ventilation. After 5 days, 15 g of treated paddy was taken in a 500 ml conical flask filled with 150 ml distilled water. Then, the flask was sealed using a stopper with silicone septum, 5 ml of HCL was injected through the syringe in the sampling adopter. The flask was kept in an ultrasonic wave-cleaner and shaken for 5 min and subsequently allowed to stand for 30 min. The headspace analysis was done using Gas Chromatography (GC-14B, Shimadzu Technologies, Columbia, USA).

2.7 Data Analysis

The experimental data were fitted to a linear equation, $Y = aX + b$, where Y is the PH₃ concentration and X is the fumigation time. The results were statistically analysed using Factorial Completely Randomized Design (FCRD) with replication at $p < 0.01$ and $p < 0.05$ level of significance through AGRES software (7.01). Fisher's least significant difference (LSD) test was followed to determine highly significant difference between the treatments and paddy moisture content. Also, probit analysis was carried out for statistically analyzing the mortality of various life stages of *T. castaneum* (LT₅₀ and LT₉₉) using polo plus software (V. 2.0).

3. Results and Discussion

3.1 Phosphine Concentration and Saturation Time

Saturation time is the extent to which maximum PH₃ concentration generated at a particular point of time. PH₃ concentration and saturation time with respect to different treatments and moisture content were depicted in Table 1. It has been observed that at 11% moisture content, maximum PH₃ concentration of 322 ppm was recorded with the corresponding saturation time of 73 h in the treatment T₁ followed by T₂ and T₄ with the PH₃ concentration of 284 ppm and 270 ppm at the saturation time of 61 h respectively. Treatment T₃ (98% CO₂ + 1 ppm PH₃) recorded lowest PH₃ concentration of 218 ppm with a saturation time of 55 h. At 14% moisture content, maximum PH₃ concentration of 340 ppm with the saturation time of 67 h was observed in T₁ followed by T₂, T₄ and T₃ (Table 1). Similarly, at 17% moisture content maximum PH₃ concentration of 351 ppm with the saturation time of 45 h was recorded. It could be explained by understanding that at higher grain moisture content elevated PH₃ concentration released in shorter period. From our studies, it can be concluded that paddy treated with 98% CO₂ + 3 ppm PH₃ was considered to be the effective treatment which enhanced PH₃ concentration due to the addition of CO₂. Comparing all the treatments, least PH₃ concentration value of 204 ppm was recorded at 11% moisture content in the treatment T₃. Several studies in the past confirmed that PH₃ concentration remains higher with the addition of CO₂ (Daolin et al., 2004). Thus, the increase in PH₃ concentration and higher grain moisture content influences the concentration at the time of fumigation. F values of PH₃ peak concentration are highly significant ($P \leq 0.01$) with respect to moisture contents, and treatments as given in the Table 2.

Table 1. Effect of different treatments (*i.e.* T₁-(98% CO₂+ 3 ppm PH₃); T₂-(98% CO₂+ 2 ppm PH₃); T₃-(98% CO₂ + 1 ppm PH₃); T₄-(3 ppm PH₃)) and grain moisture content on PH₃ generation with respect to saturation time. Values are expressed as Mean±standard deviations

Treatments	Moisture content (11% wet basis)		Moisture content (14% wet basis)		Moisture content (17% wet basis)	
	Saturation time (h)	[PH ₃] (ppm)	Saturation time (h)	[PH ₃] (ppm)	Saturation time (h)	[PH ₃] (ppm)
T ₁	73±2.31	322±0.57	67±2.31	340±1.00	45±2.31	351±1.00
T ₂	61±2.31	284±1.00	59±2.31	287±1.00	49±2.31	290±1.10
T ₃	57±2.31	204±0.74	57±2.31	235±1.00	55±2.31	218±1.53
T ₄	61±2.31	270±1.00	64±4.62	295±1.00	53±1.15	296±1.60

Table 2. Analysis of variance for PH₃ generation rate and concentration with respect to saturation time ($p < 0.01$)

Variables	df	Sum of Square	Mean Square	F
<i>PH₃</i>				
Treatment (T)*	3	52050.97	1024.11	635.65
Moisture (M)*	2	2048.22	17350.32	10769.16
TM* (Treatment × Moisture)	6	2079.77	346.62	215.14
Error	24	38.66	1.61	1.0
Total	35	56217.63	56217.63	996.96
<i>PH₃ generation rate</i>				
Treatment (T)*	3	17.78	5.92	8470.41
Moisture (M)*	2	9.22	4.61	6586.77
TM* (Treatment × Moisture)	6	5.74	0.95	1368.12
Error	24	0.01	0.0007	1.00
Total	35	32.77	0.93	1337.64

3.2 Phosphine Generation Rate

The residual concentration of the PH₃ as a function of time was fitted in linear regression to determine the PH₃ generation rate. From Figure 2, it can be concluded that at 17% moisture content, T₁ recorded highest PH₃ generation rate of 8.32 ppm/h as compared to other treatments. Similarly, at 14 and 11 % moisture content maximum PH₃ generation rate of 5.60 and 4.89 ppm/h observed in T₂ and T₁ respectively. PH₃ generation rate was lower in paddy treated with 98% CO₂ + 1 ppm PH₃. Among the three moisture levels tested, T₁ at 17% moisture content recorded highest PH₃ generation rate. This could be explained by understanding that PH₃ generates faster at higher moisture levels and the presence CO₂ enhances PH₃ toxicity due to synergistic effect. The other factors influences PH₃ gas generation includes temperature, moisture content and brand of fumigant used (Banks, 1991). Comparing to low moisture grain, high moisture grain stimulates higher PH₃ gas generation (Reed & Pan, 2000). Thus, the difference in moisture levels and treatments given to the paddy significantly ($p < 0.01$) affected the PH₃ generation rate as given in the Table 2.

3.3 Phosphine Concentration With Respect to Fumigation Time

Aluminum phosphide reacts with the moisture present in the atmosphere and produces PH₃ gas. This reaction starts slowly, then gradually increases and later declines when the aluminum phosphide is spent. Rate of PH₃ decomposition during fumigation also varies depending upon the moisture and atmospheric conditions (Figure 3). From figure, it has been observed that there was a gradual decrease in PH₃ concentration after attaining saturation point in all the treatments of paddy. Adequate moisture is required for the release of the definite toxicant or PH₃ in aluminum phosphide. It is clearly shown that the grain moisture content has direct impact on PH₃ generation. PH₃ release is enhanced at higher moisture content.

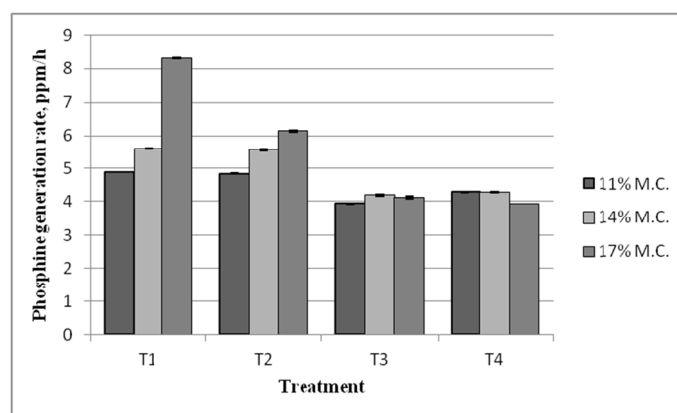


Figure 2. PH₃ generation rate in paddy fumigated with CO₂ + PH₃ at (a)11%, (b) 14% and (c) 17% moisture content (wet basis) during treatments

Note. T₁-(98% CO₂ + 3 ppm PH₃); T₂-(98% CO₂ + 2 ppm PH₃); T₃-(98% CO₂ + 1 ppm PH₃); T₄-(3 ppm PH₃).

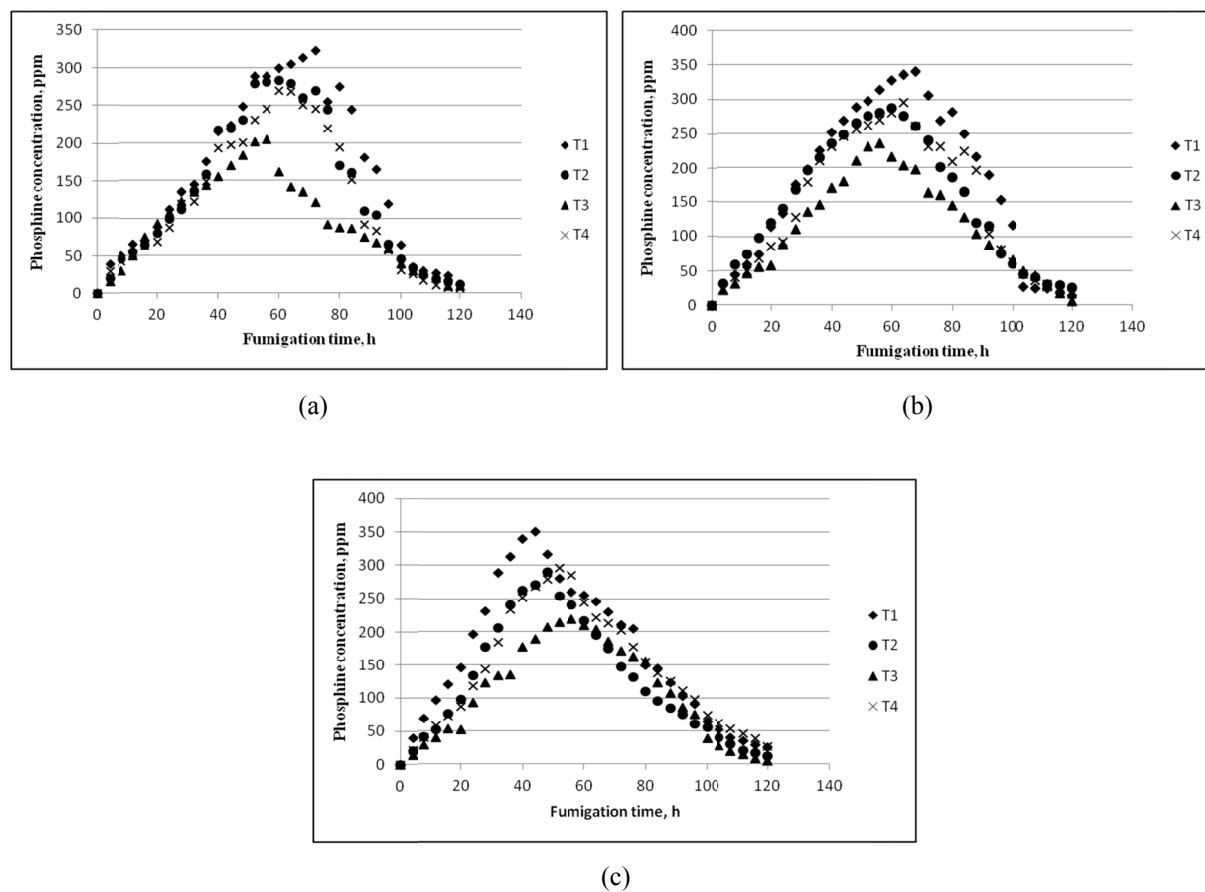


Figure 3 PH_3 concentrations in paddy fumigated with CO_2 and PH_3 at (a) 11 % (b) 14% and (c) 17 % moisture content (wet basis) during treatments

Note. T₁-(98% CO_2 + 3 ppm PH_3); T₂-(98% CO_2 + 2 ppm PH_3); T₃-(98% CO_2 + 1 ppm PH_3); T₄-(3 ppm PH_3).

3.4 Insect Mortality

The mixture of PH_3 and CO_2 significantly affects the mortality of various life stages of *T. castaneum*. The synergistic effect of CO_2 on phosphine toxicity is further supported by the fact that, CO_2 exerts lethal effects on insects causing their death by dehydration, acidification at the cellular level, and creating a lack of triglycerides for energy metabolism (Busvine, 1971; Adler, 1994; Donahaye, 1991). The LT_{99} value of *T. castaneum* in paddy at 11% moisture content was given in the Table 3. The minimum lethal time for larva was observed as 27.0 h in the treatment T₁, followed by T₂, T₄ and T₃ with the lethal time of 29.5 h, 33.7 h and 41.9 h respectively. The LT_{99} value for pupa and adult was noted as 35.5 h and 35.6 h in the treatment T₁ and T₂ respectively. LT_{50} for larva, pupa and adult were recorded as 2.9 h, 4.8 h and 1.8 h respectively. At 14% moisture content, the minimum LT_{99} value for larva was 17.1 h for the treatment T₁, followed by adult 21.7 h and pupa 36.4 h (Table 4). Similar trend was observed in paddy treated with CO_2 and PH_3 at 17% moisture content (Table 5). Among the different treatments, T₁ at 17% moisture content proved as the best treatment recorded with maximum PH_3 concentration and minimum lethal time. As the concentration of PH_3 increases time taken to achieve LT_{50} and LT_{99} decreases. Addition of CO_2 to PH_3 reduces the time and dose required for fumigation that results in quick insect mortality. Some studies have proven that the addition of CO_2 could increase the PH_3 concentration and resulted in increased toxicity levels (Donahaye & Navarro, 1989). Desmarchelier (1984) found that the mixture of phosphine and carbondioxide has synergistic effect on insect mortality as compared to either toxicant alone. The present study also confirmed that addition of CO_2 to PH_3 enhanced the insect mortality with minimum lethal time. The treatment T₁ and T₂ were significantly superior to T₃ and T₄ respectively.

Table 3. Mortality of *T. castaneum* larva, pupa and adult treated with CO₂ + PH₃ (i.e. T₁-(98% CO₂ + 3 ppm PH₃); T₂-(98% CO₂ + 2 ppm PH₃); T₃-(98% CO₂ + 1 ppm PH₃); T₄-(3 ppm PH₃)) at 11 % moisture content at 95% of confidential limit

Stage		Slope±SE	LT ₅₀ (h)	LT ₉₅ (h)	LT ₉₉ (h)	X ² (df)
Larva	T ₁	2.4±0.4	2.9(2.0-3.7)	14.1(10.3-25.3)	27.0(17.1-65.9)	4.7(16)
	T ₂	2.4±0.4	3.3(2.4-4.1)	15.6(11.3-28.1)	29.5(18.6-71.7)	8.6(16)
	T ₃	2.3±0.4	4.2(3.2-5.2)	21.4(14.7-44.0)	41.9(24.6-118.8)	10.4(16)
	T ₄	2.2±0.4	3.2(2.5-4.0)	16.9(11.9-33.4)	33.7(20.1-94.5)	8.5(16)
Pupa	T ₁	2.8±0.3	5.2(4.1-6.3)	20.3(15.3-32.2)	35.5(24.1-69.2)	35.6(25)
	T ₂	2.3±0.3	5.7(4.5-6.9)	28.9(20.6-51.3)	56.4(35.19-129.9)	28.8(25)
	T ₃	2.4±0.3	7.3(6.1-8.5)	33.7(24.7-55.2)	63.5(41.5-127.2)	24.5(25)
	T ₄	2.3±0.3	4.8(3.8-5.7)	24.3(18.3-38.3)	47.5(31.6-92.8)	13.3(25)
Adult	T ₁	1.8±0.4	2.0(0.9-2.9)	15.8(10.4-40.6)	36.6(19.3-174.5)	6.7(16)
	T ₂	2.1±0.4	2.8(1.7-3.6)	16.9(11.6-36.6)	35.6(20.3-118.5)	6.3(16)
	T ₃	2.0±0.3	3.3(2.2-4.3)	21.8(14.2-53.5)	47.2(25.2-183.1)	8.6(16)
	T ₄	1.6±0.4	1.8(0.6-2.7)	17.5(10.9-59.1)	44.7(21.2-347.9)	8.5(16)

Table 4. Mortality of *T. castaneum* larva, pupa and adult treated with CO₂ + PH₃ (i.e. T₁-(98% CO₂ + 3 ppm PH₃); T₂-(98% CO₂ + 2 ppm PH₃); T₃-(98% CO₂ + 1 ppm PH₃); T₄-(3 ppm PH₃)) at 14 % moisture content at 95% of confidential limit

Stage		Slope±SE	LT ₅₀ (h)	LT ₉₅ (h)	LT ₉₉ (h)	X ² (df)
Larva	T ₁	2.6±0.4	2.3(1.5-2.9)	9.5(7.3-15.2)	17.1(11.6-36.0)	5.2(16)
	T ₂	2.2±0.4	2.5(1.5-3.3)	13.4(9.6-25.2)	26.7(16.4-73.3)	6.4 (16)
	T ₃	2.2±0.4	3.3(2.3-4.1)	17.8(12.4-36.3)	35.8(21.1-104.5)	10.6(16)
	T ₄	2.0±0.4	2.4(1.3-3.2)	15.1(10.4-32.8)	32.3(18.4-111.9)	5.7(16)
Pupa	T ₁	2.6±0.3	4.7(3.8-5.4)	20.0(15.3-30.4)	36.4(25.0-67.0)	21.7(22)
	T ₂	2.2±0.3	5.1(4.1-6.2)	27.4(19.5-48.5)	54.7(33.9-125.4)	20.0(22)
	T ₃	2.2±0.3	6.5(5.2-8.0)	35.1(23.2-76.0)	70.3(39.7-210.0)	25.1(22)
	T ₄	2.2±0.3	4.4(3.3-5.4)	24.0(17.3-41.4)	48.3(30.3-108.7)	8.2(22)
Adult	T ₁	2.0±0.4	1.5(0.6-2.3)	10.0(7.1-20.8)	21.7(12.7-80.6)	7.2(16)
	T ₂	2.2±0.4	2.0(1.1-2.8)	11.3(8.2-21.3)	22.8(14.0-65.6)	8.6(16)
	T ₃	2.0±0.4	2.6(1.5-3.4)	16.7(11.3-37.8)	36.1(20.1-130.5)	7.6(16)
	T ₄	1.7±0.4	1.4(0.3-2.3)	12.1(7.9-41.0)	29.4(14.6-261.8)	7.9(16)

Table 5. Mortality of *T. castaneum* larva, pupa and adult treated with CO₂ + PH₃ (i.e. T₁-(98% CO₂ + 3 ppm PH₃); T₂-(98% CO₂ + 2 ppm PH₃); T₃-(98% CO₂ + 1 ppm PH₃); T₄-(3 ppm PH₃)) at 17 % moisture content at 95% of confidential limit

Stage		Slope±SE	LT ₅₀ (h)	LT ₉₅ (h)	LT ₉₉ (h)	X ² (df)
Larva	T ₁	2.2±0.5	1.3(0.3-2.0)	6.9(2.0-5.0)	13.8(8.3-58.1)	6.4(13)
	T ₂	2.0±0.5	1.3(0.4-2.1)	8.2(5.8-19.3)	17.5(10.1-84.9)	3.3(13)
	T ₃	1.9±0.5	1.6(0.6-2.5)	11.3(7.6-31.8)	25.0(13.3-147.4)	6.0(13)
	T ₄	1.5±0.4	1.1(0.09-2.1)	14.3(8.2-131.3)	40.3(16.2-2368.4)	8.0(13)
Pupa	T ₁	2.4±0.3	3.8(2.7-4.8)	18.2(13.1-33.0)	34.6(21.7-84.7)	29.4(22)
	T ₂	2.0±0.3	3.7(2.4-4.8)	24.6(16.4-54.3)	53.6(29.8-178.6)	25.5(22)
	T ₃	2.3±0.3	5.7(4.7-6.8)	29.3(20.9-51.8)	57.4(35.8-129.7)	20.4(22)
	T ₄	2.1±0.3	4.0(2.9-4.9)	23.3(16.7-41.2)	48.2(29.8-113.3)	10.5(22)
Adult	T ₁	2.2±0.5	1.6(0.7-2.3)	8.9(6.4-19.0)	18.0(10.7-67.0)	2.7(13)
	T ₂	2.2±0.5	2.0(1.0-2.7)	10.4(7.4-22.2)	20.7(12.3-71.9)	3.6(13)
	T ₃	1.9±0.4	2.4(1.2-3.3)	16.4(10.4-50.8)	36.2(18.3-213.5)	7.6(13)
	T ₄	2.1±0.4	1.9(0.9-2.7)	11.7(7.9-29.2)	24.6(13.6-112.3)	6.2(13)

4. Conclusion

From the above work it can be concluded that the use of combination of PH₃ and CO₂ is more effective than PH₃ fumigation alone. The mixture of CO₂ and PH₃ increased the PH₃ concentration and generation rate during fumigation. It has been observed that an application of 98% CO₂ with 3 ppm PH₃ and 2 ppm significantly ($p < 0.01$) improve the fumigation process by increasing PH₃ concentration and generation rate. Fumigating paddy with 98% CO₂ + 1 ppm PH₃ showed reduced effectiveness as compared to control treatment. The presence of CO₂ is also essential during fumigation which causes suffocation to insects and results in quick mortality of insects in modified atmospheric storage (Alice et al., 2015). Addition of CO₂ to PH₃ enhanced the toxicity of PH₃ and significantly reduces the LT₅₀ and LT₉₉ values. The average PH₃ concentration required for the mortality of *Tribolium castaneum* (adult, pupa and larva) at different grain paddy moisture content was determined. However, egg mortality has to be studied for the above mentioned treatments to prevent reinfestation of insects during storage. PH₃ residues in all the treatments including control were recorded below the recommended level (0.1 ppm). Hence it is considered as safe for consumption in all the treatments including control.

Acknowledgements

The authors gratefully acknowledge Department of Science & Technology (DST), Science and Engineering Research Board (SERB) and Ministry of Food Processing Industries (MoFPI) for funding the scheme and Director, IIFPT for the support and guidance.

References

- Alice, R. P., Sujeetha, J., Meenatchi, R., Venkatesan, P., & Brimapureeswaran, R. (2015). Nitrogen based modified atmosphere for the management of Cigarette beetle, *Lasioderma serricorne* in turmeric. *International Journal for Science and Advance Research in Technology*, 1(11), 84-87.
- Banks, H. J. (1991). Influence of water and temperature on release of phosphine from aluminium phosphide-containing formulations. *Journal of Stored Products Research*, 27, 41-56. [https://doi.org/10.1016/0022-474X\(91\)90031-7](https://doi.org/10.1016/0022-474X(91)90031-7)
- Daolin, G., Xiaoping, Y., Zhanggui, Q., Longde, Y., Zili, Y., & Tianke, J. (2004). Using carbon dioxide to enhance the efficacy of phosphine fumigation. In E. J. Donahaye, S. Navarro, C. Bell, D. Jayas, R. Noyes, & T. W. Phillips (Eds.), *Proc. Int. Conf. Controlled Atmosphere and Fumigation in Stored Products* (pp. 485-492). Gold-Coast Australia, FTIC Ltd. Publishing, Israel.
- Desmarchelier, J. M. (1984). *Effect of carbon dioxide on the efficacy of phosphine against different stored product pests* (No. 220, p. 57). Federal Biological Research Centre for Agriculture and Forestry, Berlin.
- Donahaye, E., & Navarro, S. (1989). Sensitivity of two dried fruit pests to methyl bromide alone, and in combination with carbon dioxide or under reduced pressure. *Tropical Science*, 29, 9-14.
- Hasan, M. M., Aikins, M. J., Schilling, W., & Phillips, T. W. (2016). Efficacy of controlled atmosphere treatments to manage arthropod pests of dry-cured hams. *Insects*, 7(3), 44. <https://doi.org/10.3390/insects7030044>
- Leelaja, B. C., Rajashekar, Y., Reddy, P. V., Begum, K., & Rajendran, S. (2007). Enhanced fumigant toxicity of allyl acetate to stored-product beetles in the presence of carbon dioxide. *Journal of Stored Products Research*, 43, 45-48. <https://doi.org/10.1016/j.jspr.2005.09.004>
- Leesch, J. G. (1992). Carbon dioxide on the penetration and distribution of phosphine through wheat. *J Econ. Entomol*, 85, 157-161. <https://doi.org/10.1093/jee/85.1.157>
- Mau, Y. S., Collins, P. J., Darglish, G. J., Nayak, M. K., & Ebert, P. R. (2012). The rph2 gene is responsible for high level resistance to phosphine in independent field strains of *Rhyzopertha dominica*. *PLoS ONE*, 7, e34027. <https://doi.org/10.1371/journal.pone.0034027>
- Meenatchi, R., & Alagusundaram, K. (2014). The current status of fumigation in India: Constrains and recent developments. *Trends in Entomology*, 10, 97-103.
- Meenatchi, R., Brimapureeswaran, R., Alagusundaram, K., & Sujeetha, A. R. P. (2015). Design and development of carbon dioxide fumigation setup with automatic gas controlling system for safe storage of grains. *International Journal of Emerging Technology and Innovative Engineering*, 1(6), 29-32.
- Rajendran, S. (2001). Alternatives to methyl bromide as fumigants for stored food commodities. *Pesticide Outlook*, 12(6), 249-253. <https://doi.org/10.1039/B110550G>

- Reed, C., & Pan, H. (2000). Loss of phosphine from unsealed bins of wheat at six combinations of grain temperature and grain moisture content. *Journal of Stored Products Research*, 36, 263-279. [https://doi.org/10.1016/S0022-474X\(99\)00049-1](https://doi.org/10.1016/S0022-474X(99)00049-1)
- Ren, Y. L., O'Brien, I. G., & Whittle, G. P. (1994). Studies on the effect of carbon dioxide in insect treatment with phosphine. In E. Highley, E. J. Wright, H. J. Banks, & B. R. Champ (Eds.), *Proc. of the 6th Int. Working Conf. on Stored-product Protection, 17-23 April 1994* (pp. 173-177). Canberra, CAB, Wallingford, UK.
- Riudavets, J., Castañé, C., Alomar, O., Pons, M. J., & Gabarra, R. (2010). The use of carbon dioxide at high pressure to control nine stored-product pests. *Journal of Stored Products Research*, 46, 228-233. <https://doi.org/10.1016/j.jspr.2010.05.005>
- Tumaming, J., Depalo, M., Garnier, J. P., & Mallari, R. (2012). ECO₂ Fume and Vaporph₃ OS phosphine fumigants—Global application updates. In S. Navarro, H. J. Banks, D. S. Jayas, C. H. Bell, R. T. Noyes, A. G. Ferzli, M. Emekci, A. A. Isikber, & K. Alagusundaram (Eds.), *Proc. of the 9th Int. Conf. on Controlled Atmosphere and Fumigation in Stored Products, 15-19 October 2012* (pp. 363-373). Antalya, Turkey, ARBER Professional Congress Services, Turkey.
- Valmas, N., & Ebert, P. R. (2006). Comparative toxicity of fumigants and a phosphine synergist using a novel containment chamber for the safe generation of concentrated phosphine gas. *PLoS ONE*, 1, e130. <https://doi.org/10.1371/journal.pone.0000130>
- Williams, P. (1985). Toxicity of methyl bromide in carbon dioxide enriched atmospheres to beetles attacking stored grain. *General Applied Entomology*, 17, 17-24.

Copyrights

Copyright for this article is retained by the author(s), with first publication rights granted to the journal.

This is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/4.0/>).